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FUJIFILM Cellular Dynamics

Quick Guide

iCell® Pericytes, 01279

Handling and Storage

Upon receipt, immediately transfer components to the proper storage temperature.

Component (Catalog #) – Amount	Storage Temperature
iCell® Pericytes, 01279 (C1241) – 1 ml cryovial	Vapor Phase of Liquid Nitrogen
iCell® Astrocyte & Pericyte Medium (M1041) – 30 ml	-20°C
iCell® Plating Supplement A (M1057) – 0.2 ml	-20°C

Preparing Cell Culture Surfaces

The recommended plate type for cell culture of iCell® Pericytes is a standard tissue culture-treated plastic plate. For imaging and staining, best results are obtained with Poly-D-Lysine (PDL) pre-coated plates.

Preparing Thawing Medium

1. Thaw iCell® Astrocyte & Pericyte Medium overnight at 4ºC prior to use.
2. Determine how much Thawing Medium is necessary for the initial thaw and plating steps. Only prepare that volume.
3. Combine **2 µl of iCell Plating Supplement A per 1 ml of iCell Astrocyte & Pericyte Medium**.
4. Store remaining iCell Astrocyte & Pericyte Medium at 4ºC for subsequent cell culture. Always equilibrate medium to room temperature prior to use.

Thawing iCell Pericytes

Avoid vigorous shaking or vortexing of the cells during thaw.

1. Transfer 3 ml of Thawing Medium to a 15 ml conical centrifuge tube.
2. Thaw the cryovial of iCell Pericytes in a 37°C water bath for 2 minutes.
3. Spray the cryovial with 70% ethanol.
4. Slowly transfer the contents to the 15 ml tube with a pipette.
5. Rinse the cryovial with 1 ml of Thawing Medium and slowly transfer the rinse to the 15 ml centrifuge tube.
6. Centrifuge the cell suspension in the 15 ml tube at 600 × *g* for 6 minutes.
7. Aspirate the supernatant, careful not to disturb the cell pellet.
8. Obtain the number of viable cells/vial from the Certificate of Analysis (CoA).
9. Adjust the final cell density by resuspending the cell pellet in the appropriate volume of Thawing Medium.

Note: *For PDL-treated 96-well plates, a seeding density of 25,000 cells/cm² is recommended. For larger vessel formats, a slightly lower seeding density of 20,000 cells/cm² is acceptable. Optimal plating density may vary by application. See Table 1 for recommendations.*

Table 1: Recommended Cell Plating Information (per well)

Cell Culture Vessel	Surface Area	Volume	Cell Number	Cell Density (viable cells/ml)
6-well Plate	9.6 cm²	2 ml	192,000	96,000
12-well Plate	3.8 cm²	1 ml	76,000	76,000
96-well PDL-coated Plate	0.32 cm²	0.2 ml	8,000	40,000

Plating and Maintaining the Cells

1. Gently pipette the cell suspension two times to mix.
2. Dispense iCell Pericytes into the cell culture vessel.
3. Culture the cells in an incubator maintained at 37°C, 5% CO_2.
   Perform a full medium exchange every other day using the volumes listed in Table 1.

*Do **NOT** passage iCell Pericytes. Use the cells for co-culture experiments within 4-5 days post-thaw.*

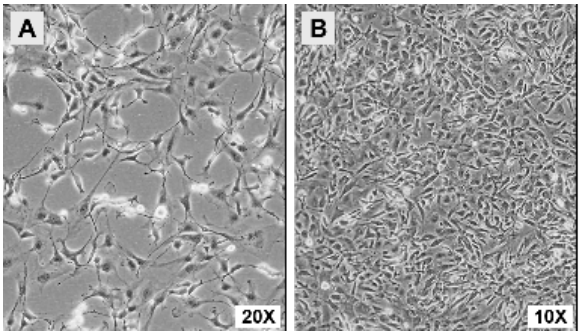


Figure 1. iCell® Pericytes, 01279 plated at 20k viable cells/cm² and imaged on **(A)** Day 1 and **(B)** Day 5 post-plating.

Table 2: Required Consumables

Component	Vendor	Catalog #
Culture Vessels, TC Treated, Sterile	Multiple Vendors	NA
96w PDL Pre-coated Plates, Sterile	Multiple Vendors	NA
TrypLE™ Select Enzyme (1X)	Gibco	12563-011
D-PBS, no calcium, no magnesium	Gibco	14190-094

Table 3: Companion iCell® Products

Product	Catalog #
iCell® Astrocytes 2.0, 01279	C1249
iCell® Brain Microvascular Endothelial Cells, 01279	C1239

Note: *iCell Pericytes can be co-cultured with iCell Astrocytes 2.0 and/or iCell Brain Microvascular Endothelial Cells (BMEC) directly from thaw.*

Table 4: Co-Culture Application Protocols [1]

Document	ID #
iCell BBB Isogenic Kit User's Guide	X1045
Adapting the iCell BBB Kit TEER assay to 96-well Format	X4024

[1] *Available for download at www.fujifilmcdi.com.*

Contacting Technical Support

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