

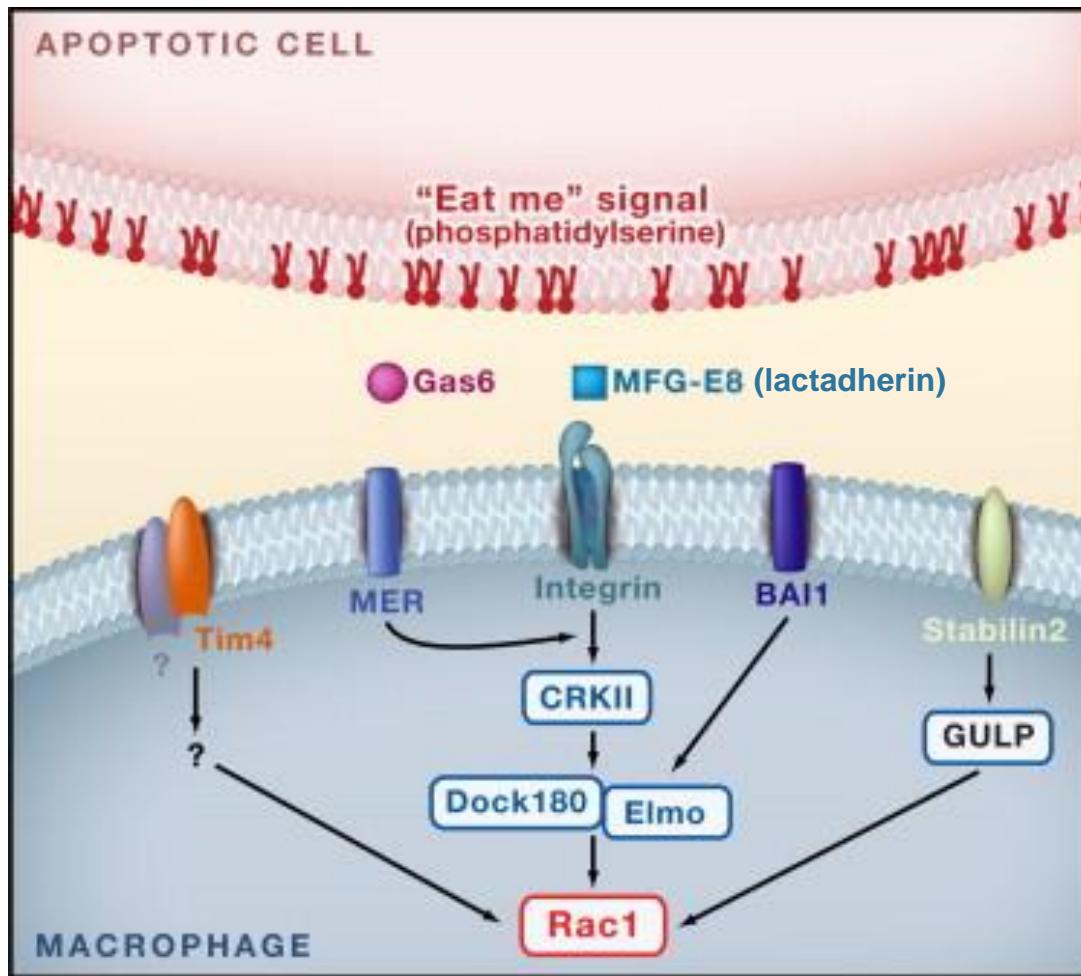
TIM4-affinity methods targeting phosphatidylserine for isolation or detection of extracellular vesicles (PS-affinity methods)

Rikinari HANAYAMA
WPI Nano Life Science Institute
KANAZAWA UNIVERSITY



ISEV2022
Platinum Sponsored Session
Fujifilm-Wako Seminar

Identification of phosphatidylserine receptors



Nagata S, Hanayama R et al., Cell. (2010)

Miyanishi et al., Nature (2007)

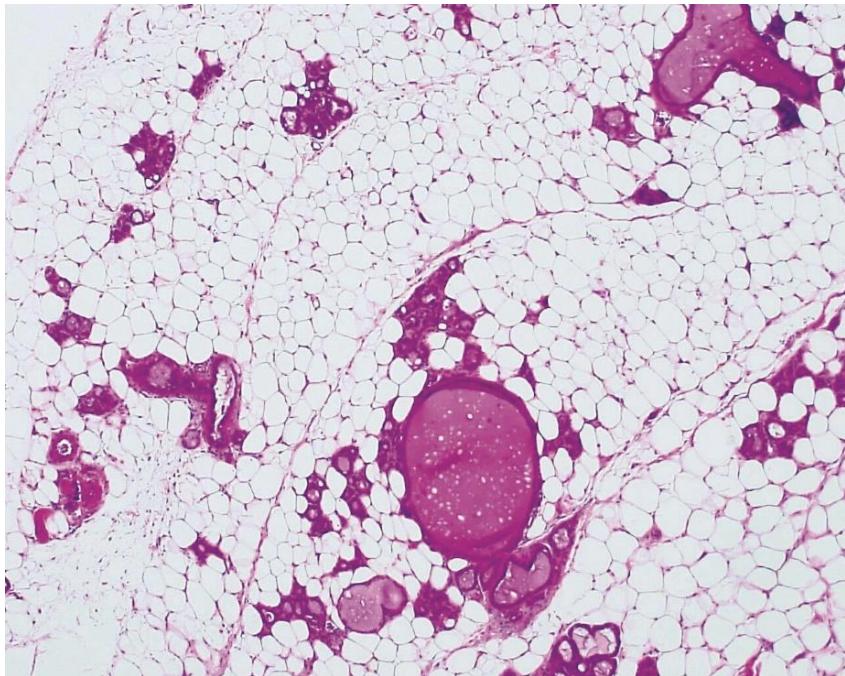
Hanayama et al., Science (2004)

Hanayama et al., Nature (2002)

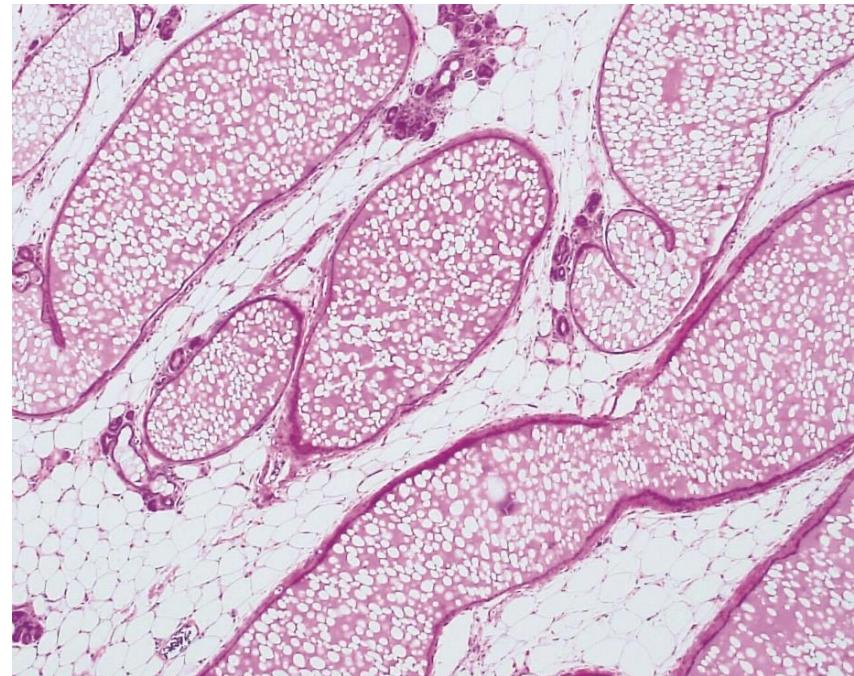
Milk fat globules are cleared by MFG-E8

Upon weaning (lactation ends),
the developed mammary gland undergo involution

WT



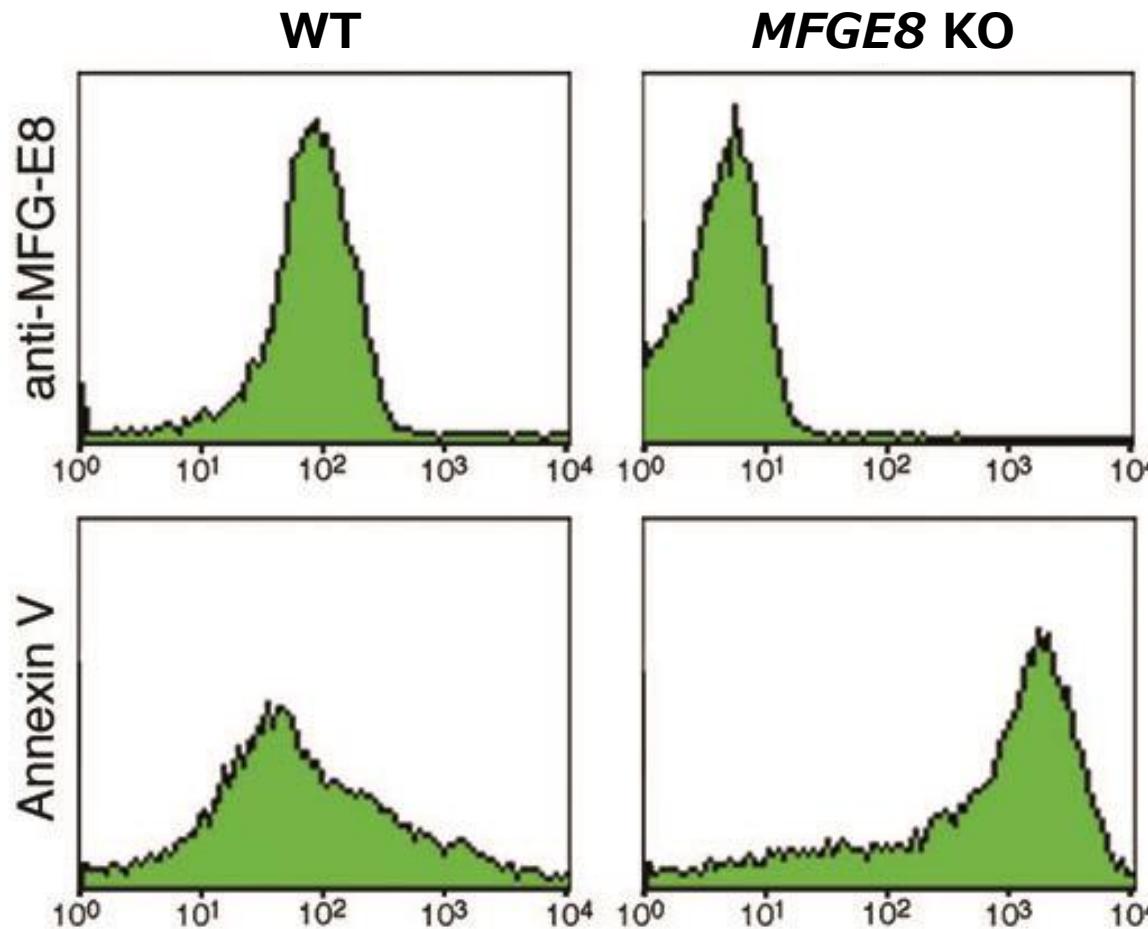
MFGE8 KO



Hanayama et al.,
PNAS (2005)

Milk fat globules expose phosphatidylserine (PS)

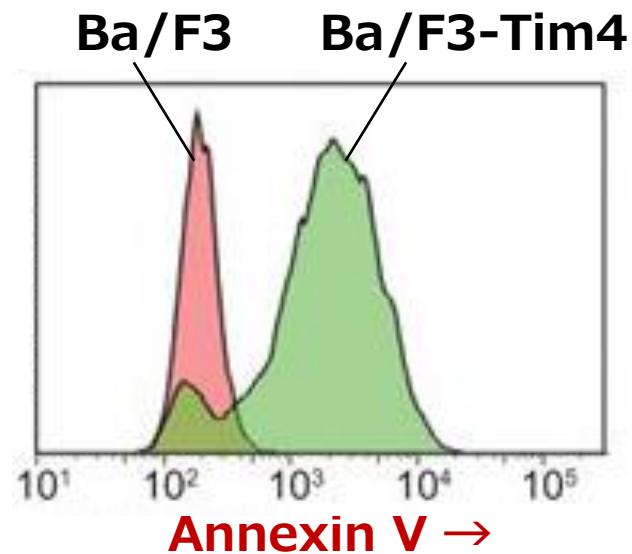
Flow cytometric analysis of single vesicles



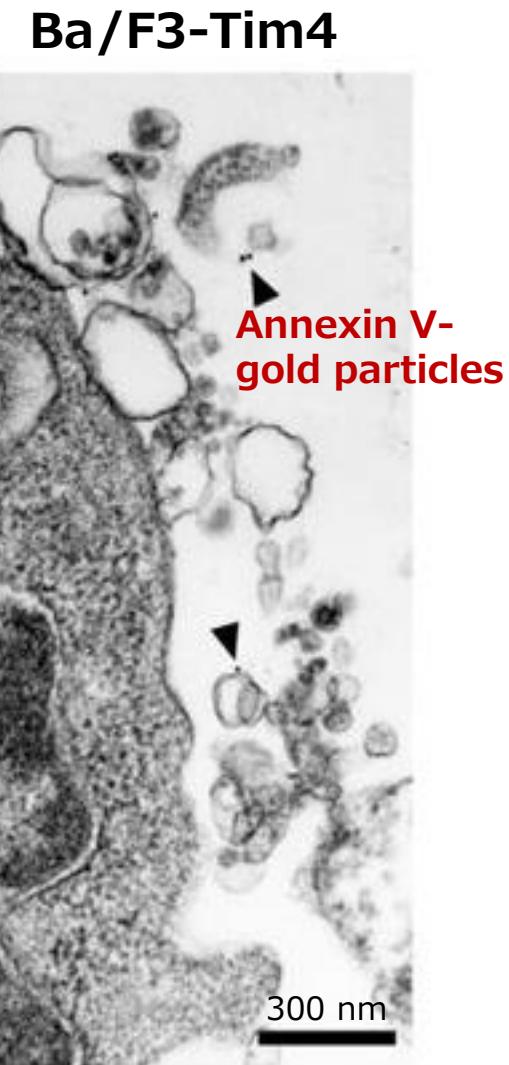
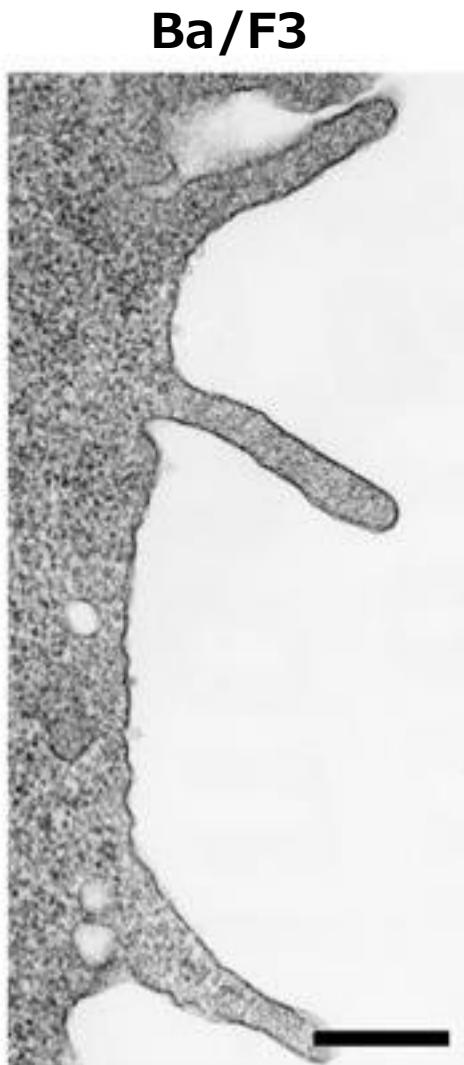
Masking of phosphatidylserine by MFG-E8

Hanayama et al.,
PNAS (2005)

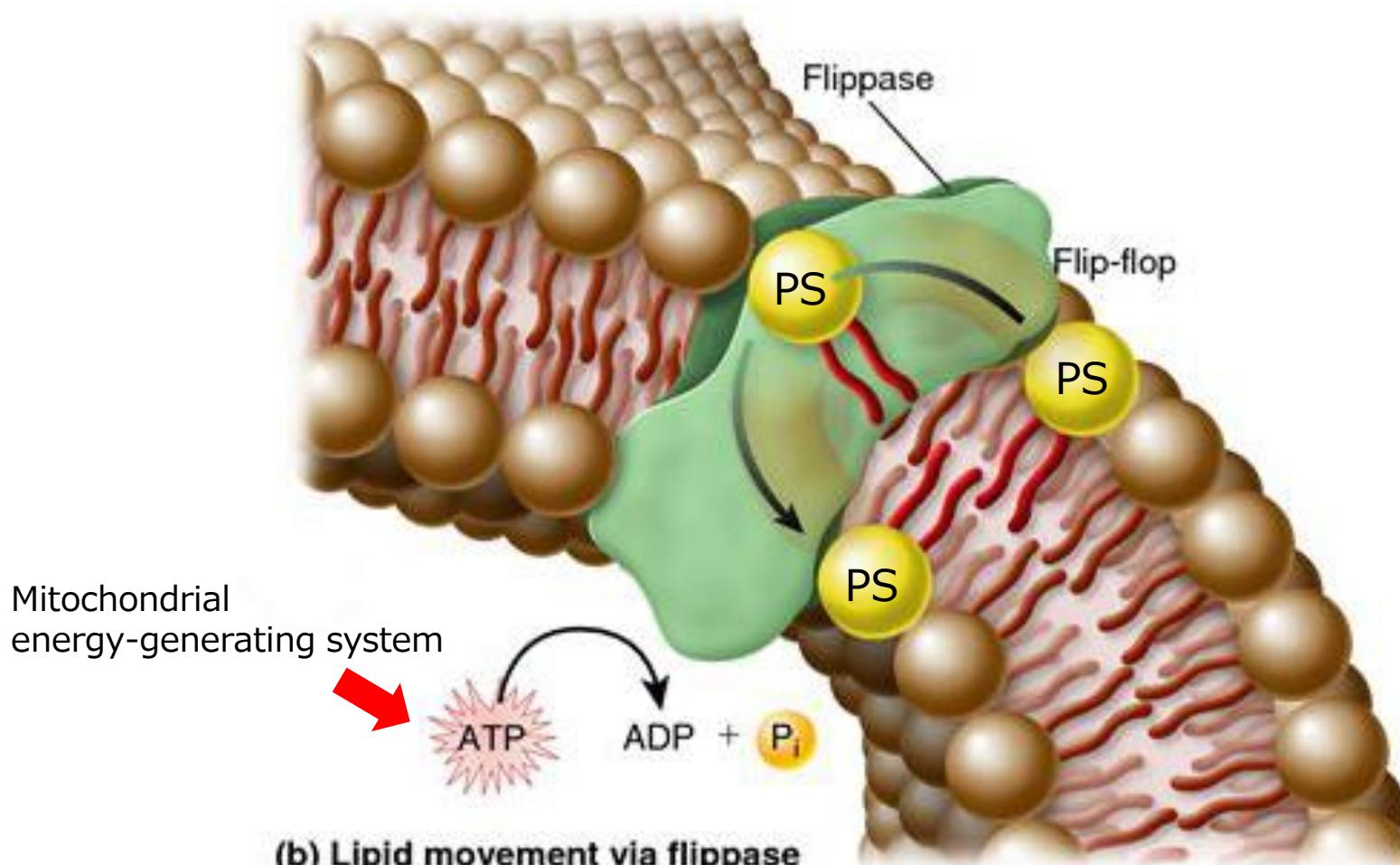
Tim4-associated EVs expose phosphatidylserine (PS)



Miyanishi et al., Nature (2007)



Exhaustion of ATP might be cause of PS exposure



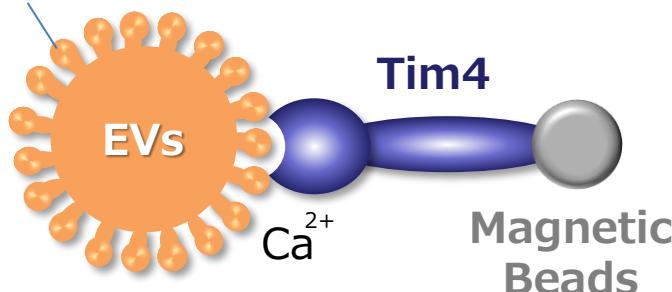
Segawa et al., Science (2014)

From Biology Forum Gallery

**A high purity isolation method of EVs
using Tim4**

A novel affinity-based EV isolation method

Phosphatidylserine (PS)



Release

Nakai & Hanayama,
Scientific Reports (2015)

FUJIFILM Wako



Life Science

Exosome isolation by novel affinity molecule
MagCapture™

Exosome Isolation Kit PS

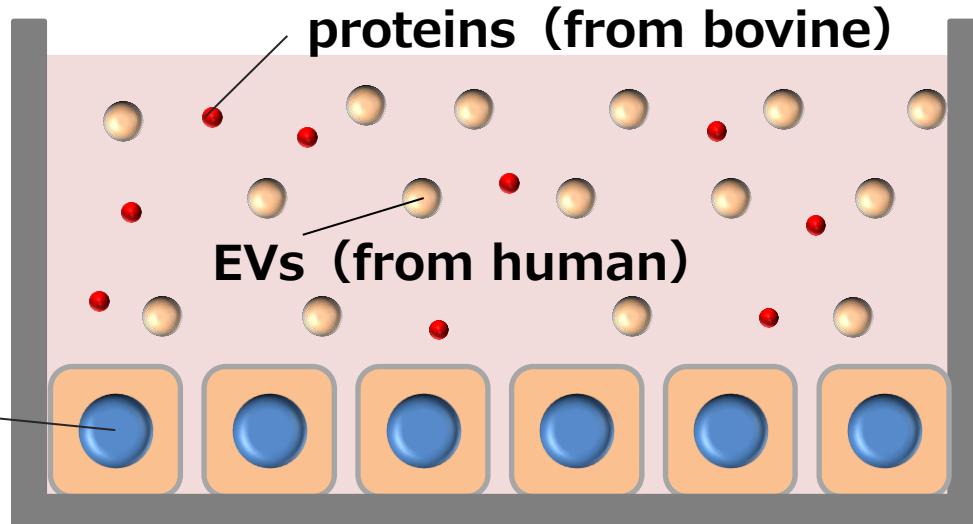
Affinity method for phosphatidylserine (PS) on membrane surface of extracellular microvesicles



Isolation of small EVs from human cells

Cultured in medium containing **5% FBS**
(EV-depleted by UC and PEG prep)

Human cells
K562 cells



The conditioned medium

↓ 300 × g, 10min
↓ 2,000 × g, 20min
↓ 10,000 × g, 30min
↓ **10K supernatants**, 0.22um filtration

sEVs



① Tim4 method

MagCapture
Exosome Isolation Kit PS

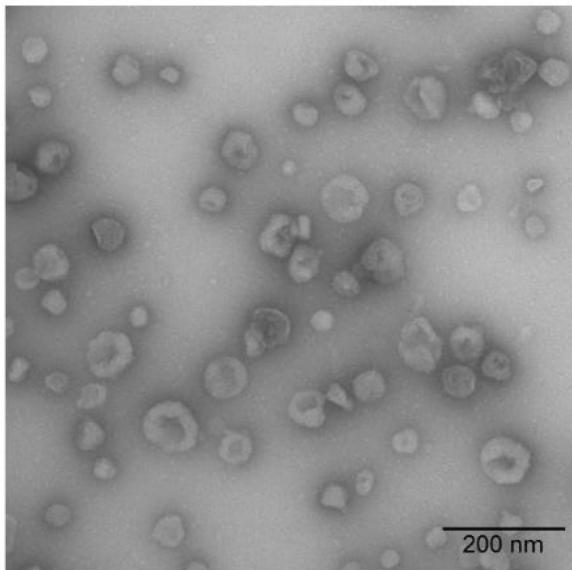
② Ultracentrifugation

↓ 100,000 × g, 120min
↓ PBS wash
↓ 100,000 × g, 120min

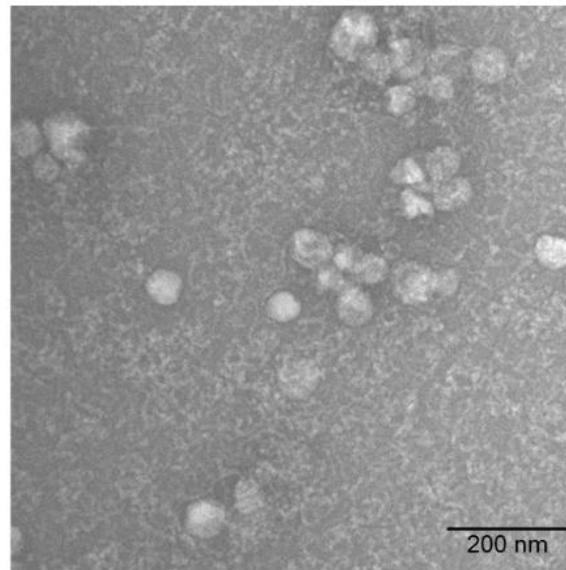
TEM and NTA analyses of isolated sEVs

a

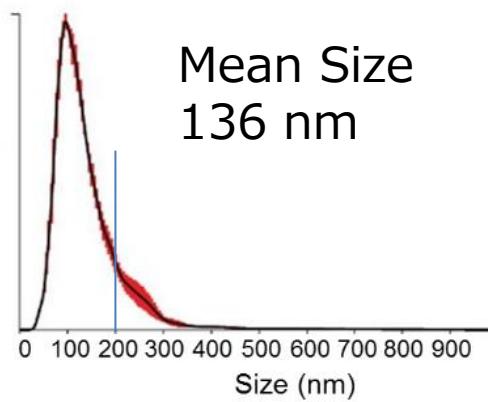
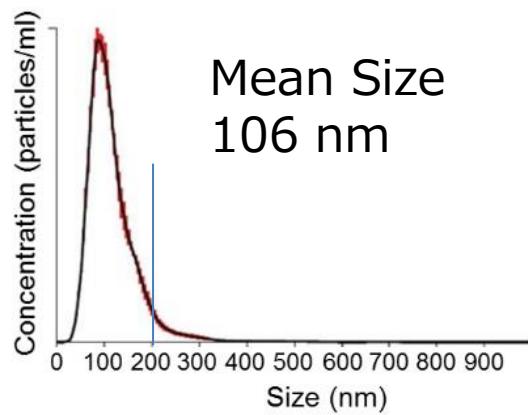
Tim4 method



Ultracentrifugation



b

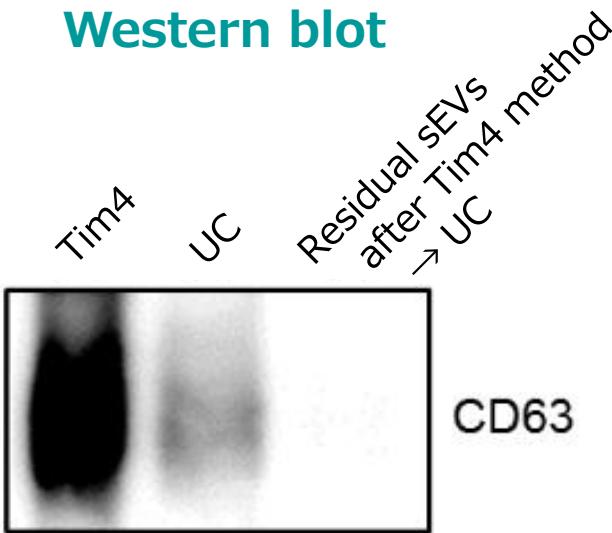


Comparison of the purity of sEVs

sEVs from K562

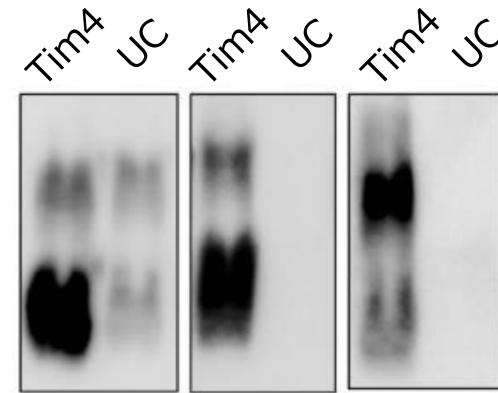
(100 ng/lane)

Western blot

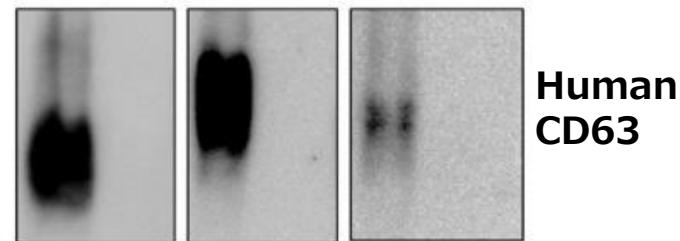


sEVs from various cell types

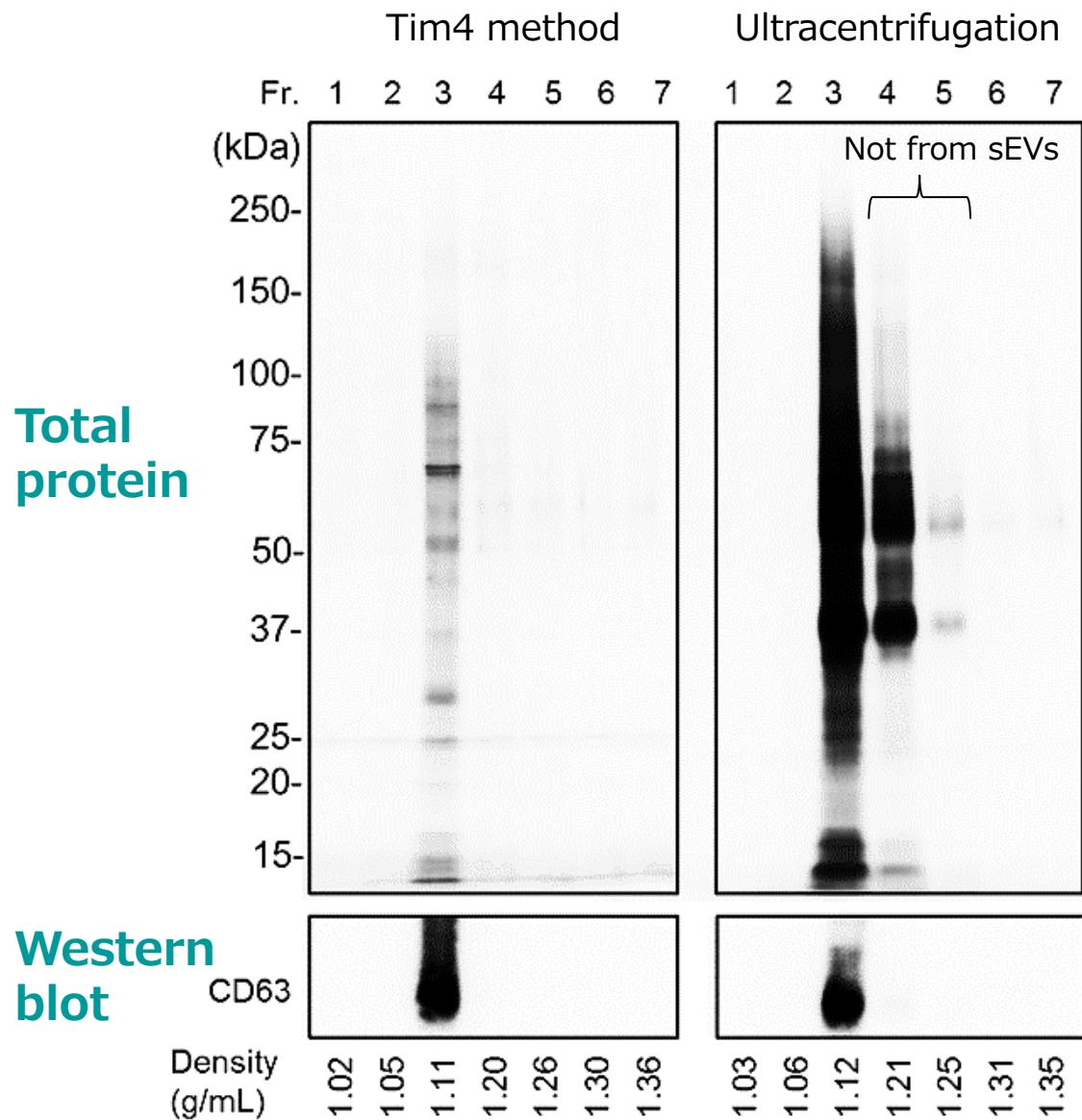
B16F10 RAW264 Adipocyte



293T PC-9 HeLa



Density gradient fractionation



Top 15 proteins identified in the isolated sEVs

White columns: human proteins (from sEVs)
Gray columns: bovine proteins (from FBS)

	Tim4 method	Ultracentrifugation
1	Heat shock cognate 71 kDa protein	DNA-PK catalytic subunit
2	Annexin A6	Transferrin receptor protein 1
3	Transferrin receptor protein 1	Serum albumin
4	V-type proton ATPase subunit A	ATP-dependent RNA helicase A
5	Flotillin-2	Tubulin beta-5 chain
6	Programmed cell death 6	Heat shock cognate 71 kDa protein
7	4F2 cell-surface antigen heavy chain	Fatty acid synthase
8	Annexin A1	4F2 cell-surface antigen heavy chain
9	Kinase D-interacting substrate	U5 small nuclear RNP helicase
10	Annexin A2	Tubulin beta-4B chain
11	Flotillin-1	Ribonucleoprotein M
12	V-type proton ATPase subunit B	Hemoglobin fetal subunit beta
13	Annexin A11	Clathrin heavy chain 1
14	Annexin A7	Fibronectin
15	Syntenin-1	Tubulin alpha-1B chain

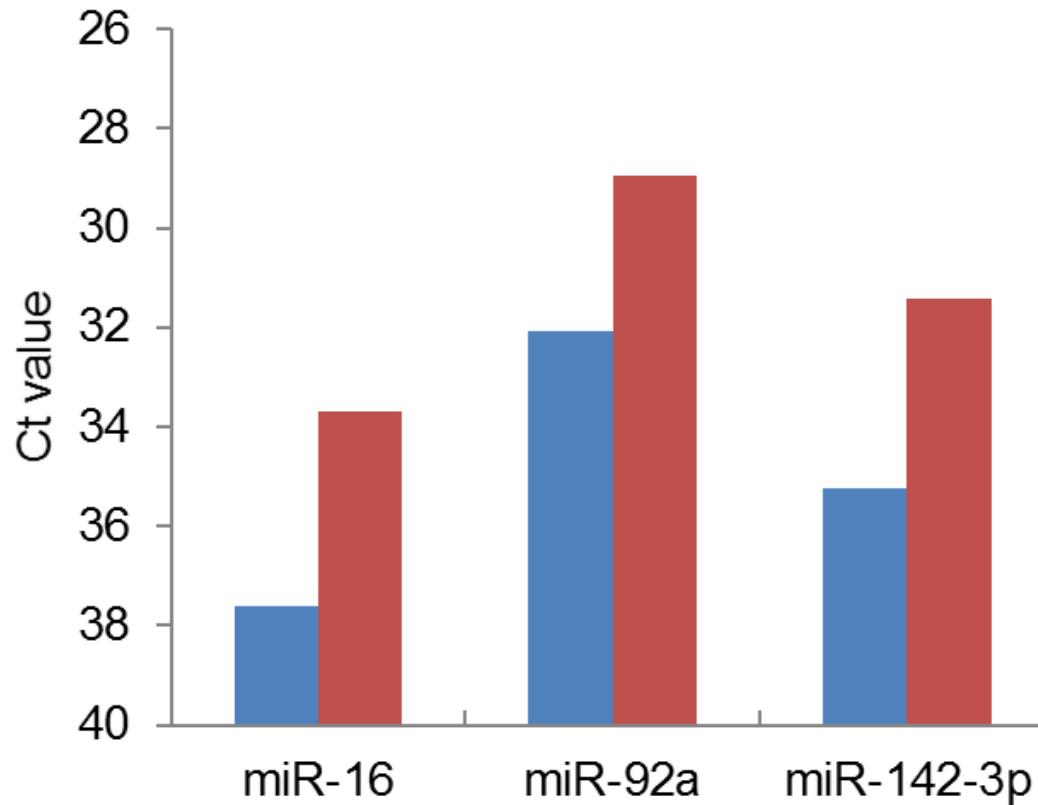
Known sEV markers

Nuclear proteins

Comparison of enrichment of sEV microRNAs

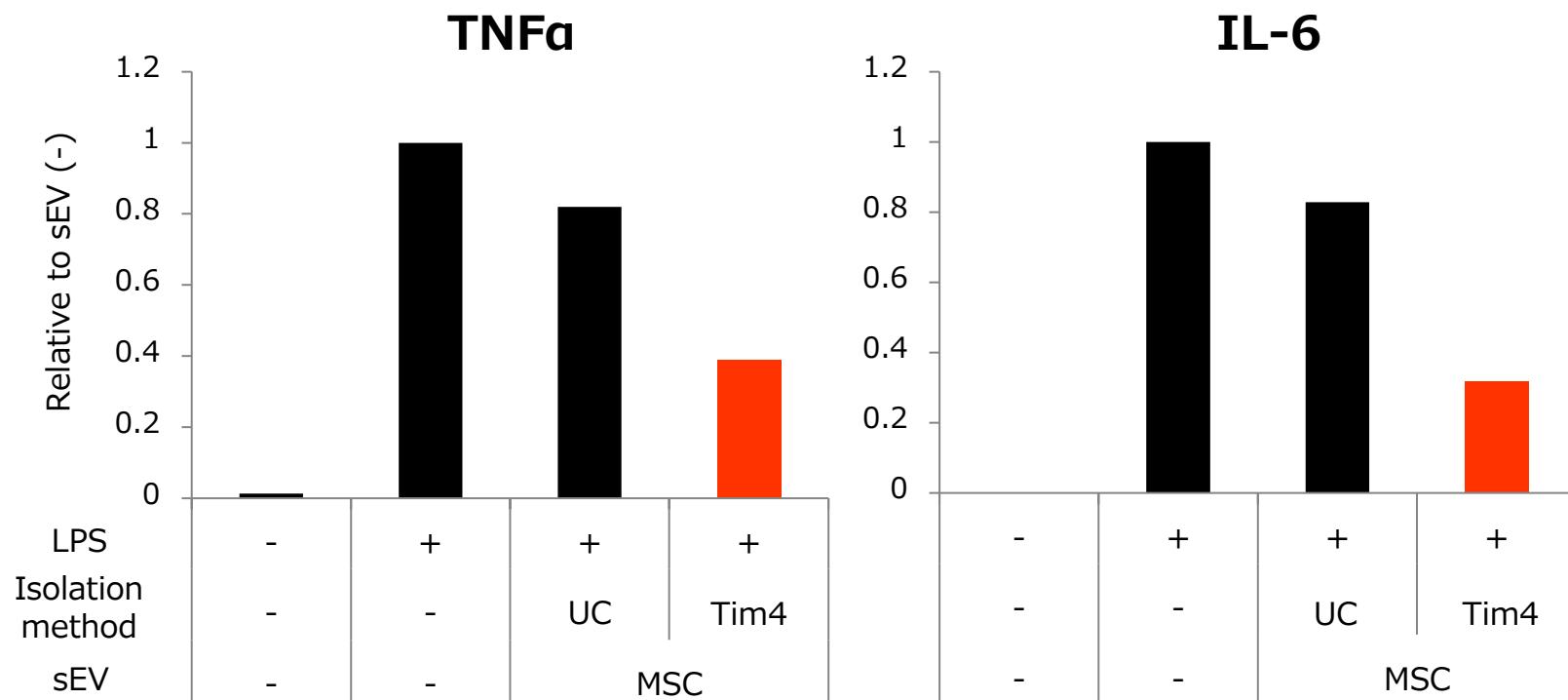
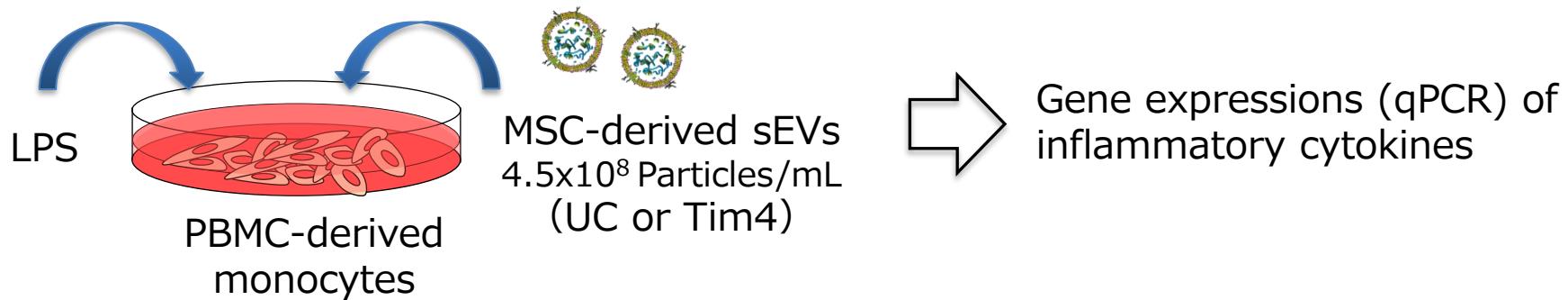
qPCR analyses in the isolated sEV fractions

■ Tim4 method ■ Ultracentrifugation

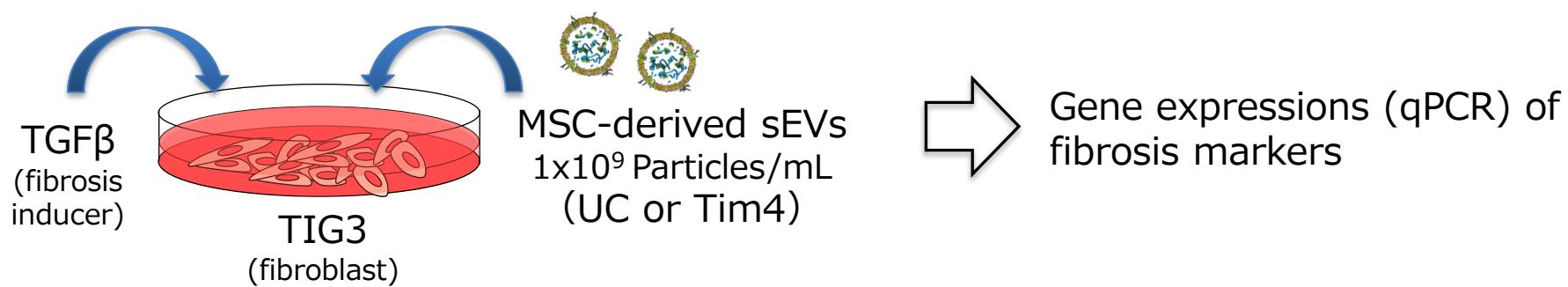


10 times more sEV RNAs were isolated by Tim4 method

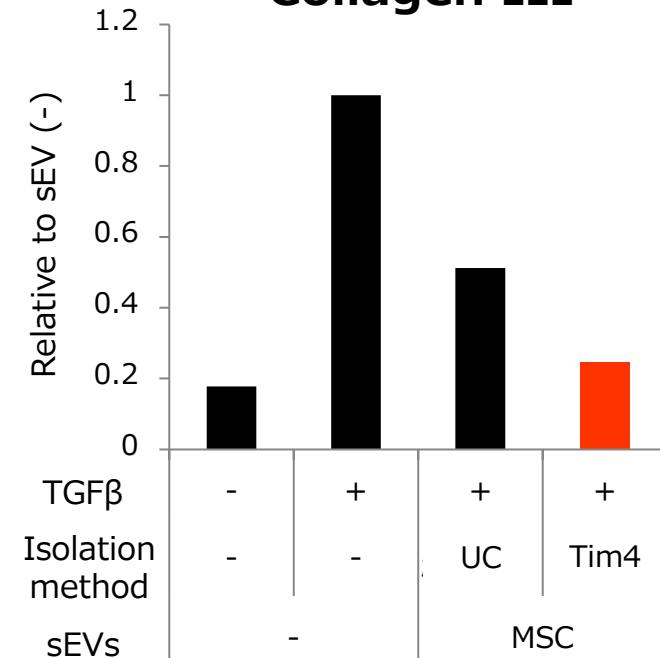
Comparison of anti-inflammatory activity of sEVs



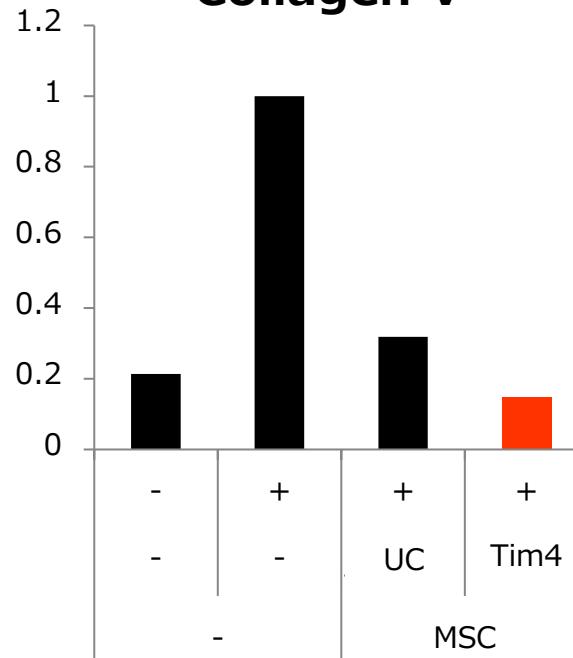
Comparison of anti-fibrotic activity of sEVs



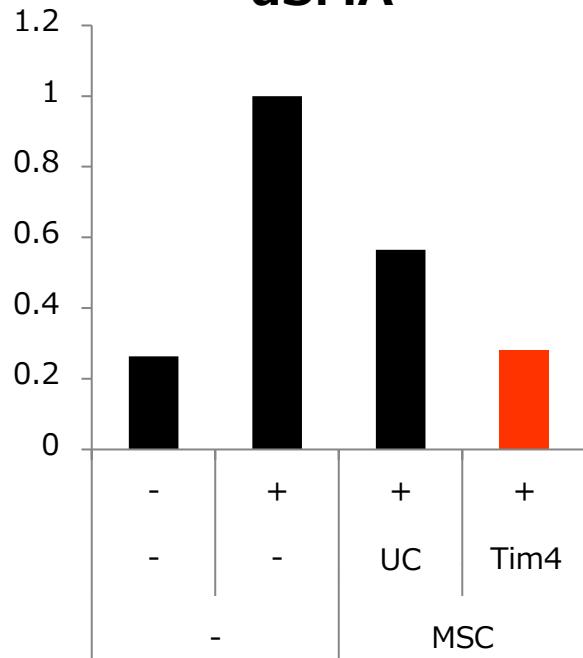
Collagen III



Collagen V



aSMA



What % of sEVs expose PS & can be isolated by Tim4?

Luc-Lamp2 expressing sEVs



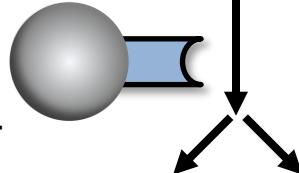
PS(+) -sEVs



PS(-) -sEVs

Matsumoto et al., iScience (2021)

Tim4-beads



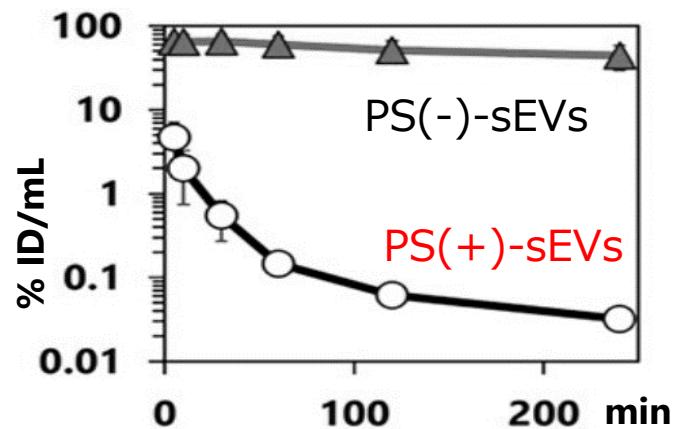
Captured fraction

88.4%

Non-captured fraction

11.6%

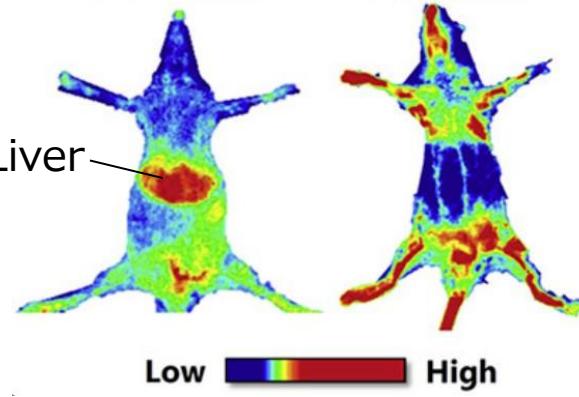
Clearance in serum after iv injection



PS(+) -sEVs

PS(-) -sEVs

Liver

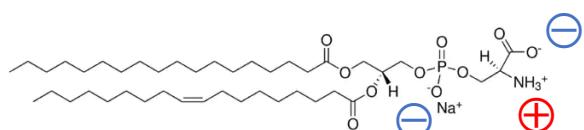


Mass balance (% of Luc recovered rate)

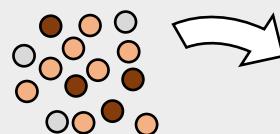
For EV-based DDS

Which subpopulations could be enriched?

Phosphatidylserine

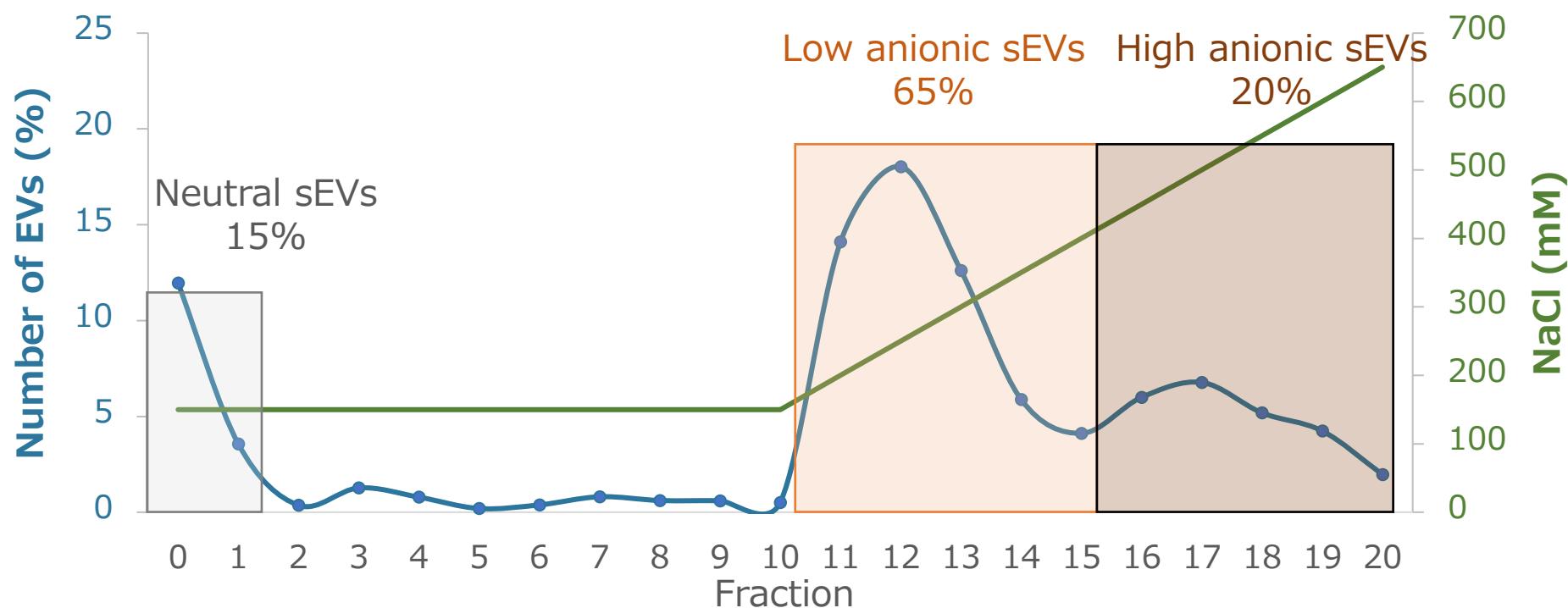


Bulk sEVs (isolated by UC)

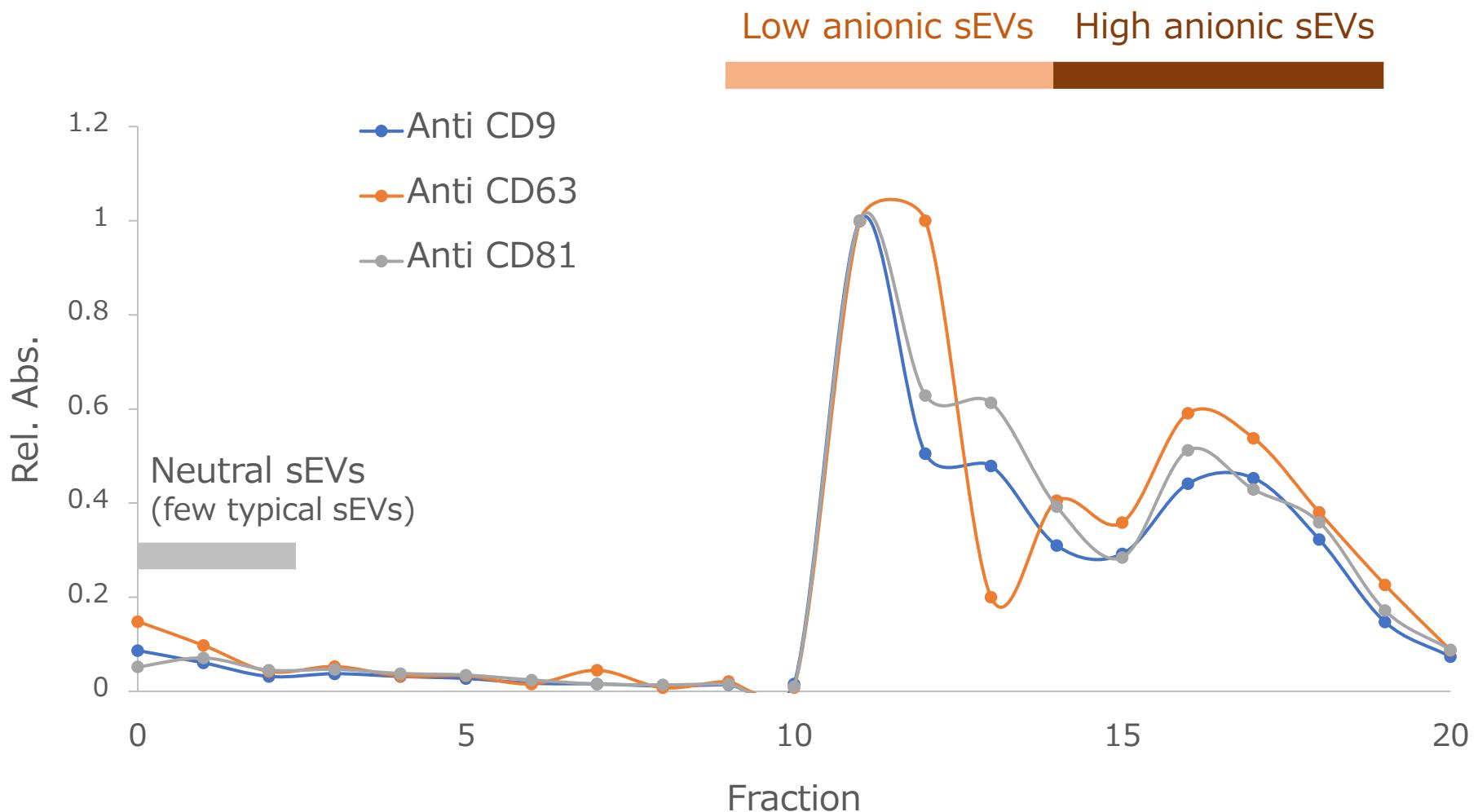


High anionic sEVs
Low anionic sEVs
Neutral sEVs

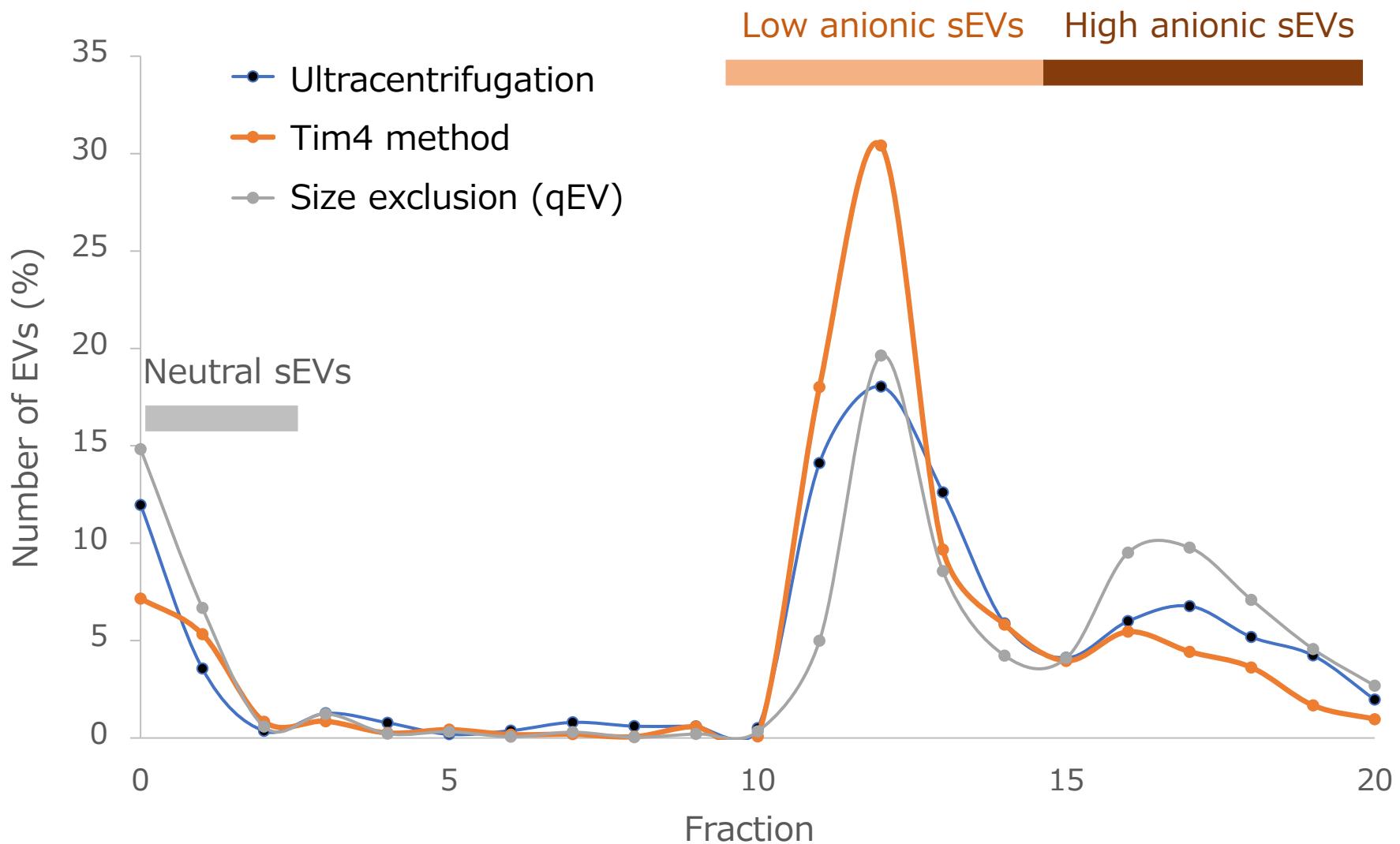
Anion exchange column
(DEAE-Sepharose)



Anti-tetraspanin Abs binding to each fraction

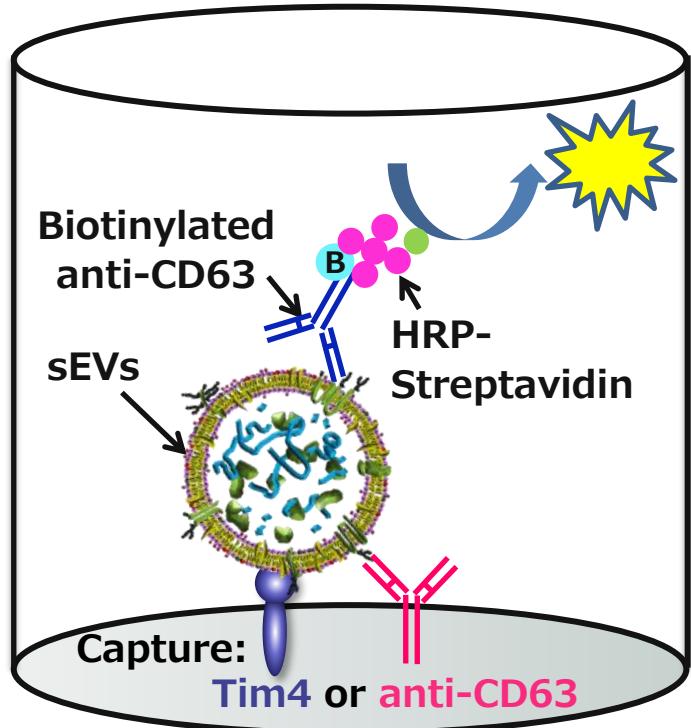


Comparison of the number of sEV subpopulations

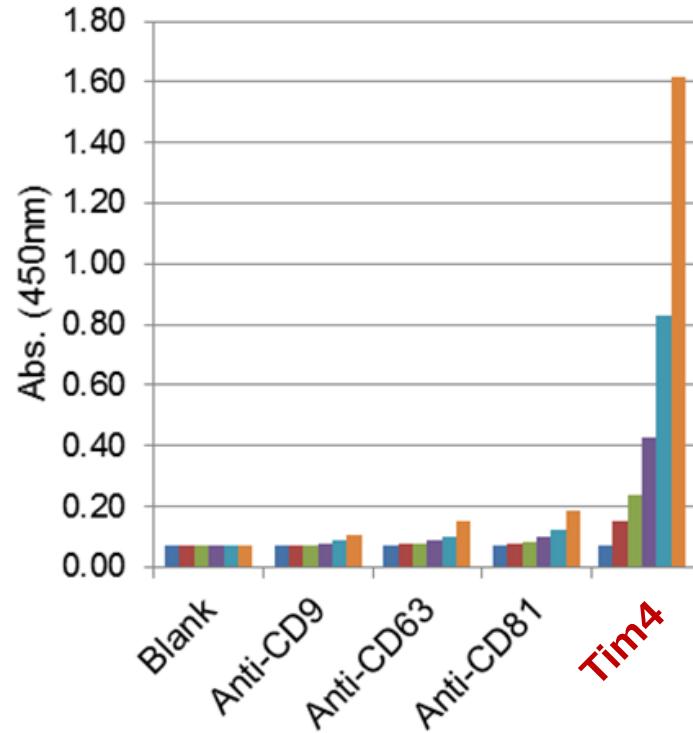


Sensitive quantification of EVs by ELISA using Tim4

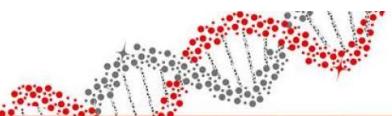
Sensitive quantification of EVs by Tim4-ELISA



1000 times more sensitive
(limit: 16ng vs 11pg)



FUJIFILM Wako



A novel tool for qualitative and quantitative analysis of extracellular vesicles
in sample of cell culture supernatant

PS Capture™

Exosome ELISA Kit (Streptavidin HRP)

Proteins coated on a microtiter plate

Concentrations
of sEVs

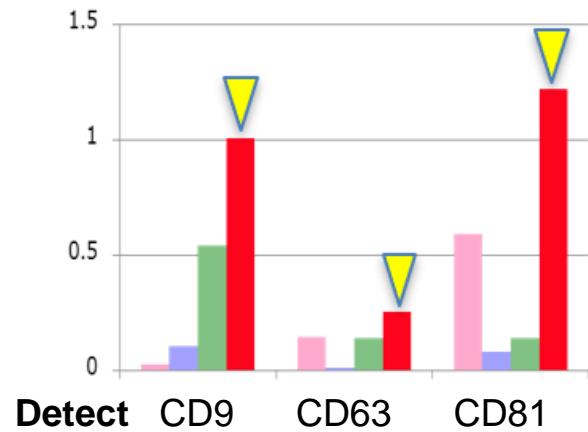


0 ← 2µg/ml (1/2 dilution)

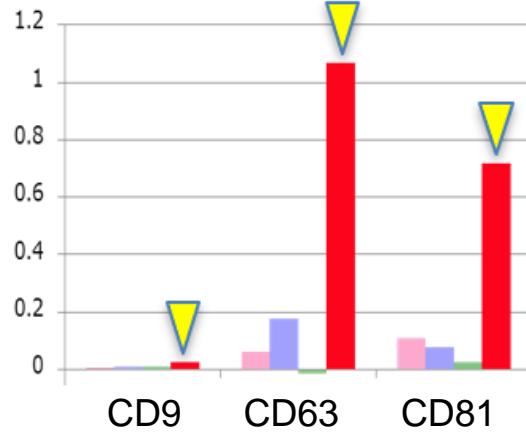
Sensitive quantification of sEVs from various cell types

Capture : CD9 Ab CD63 Ab CD81 Ab Tim4

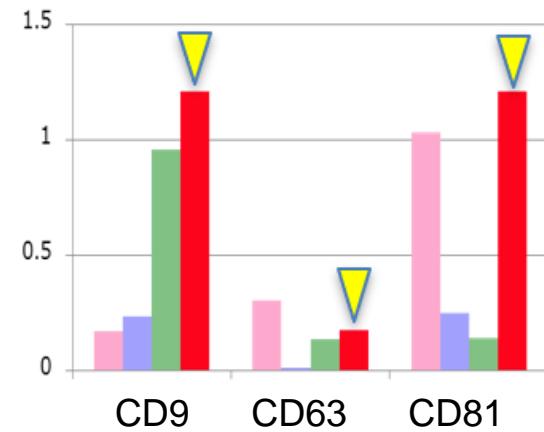
iPS



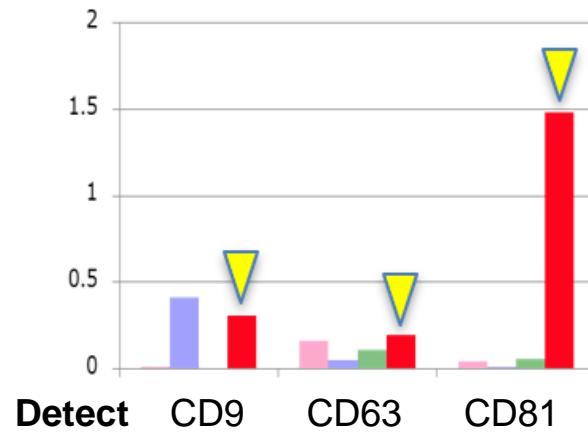
K562



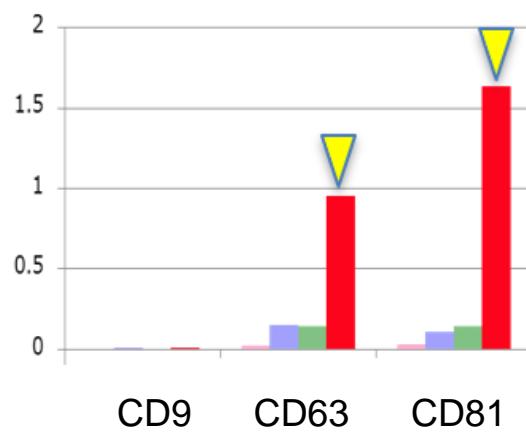
LNCaP



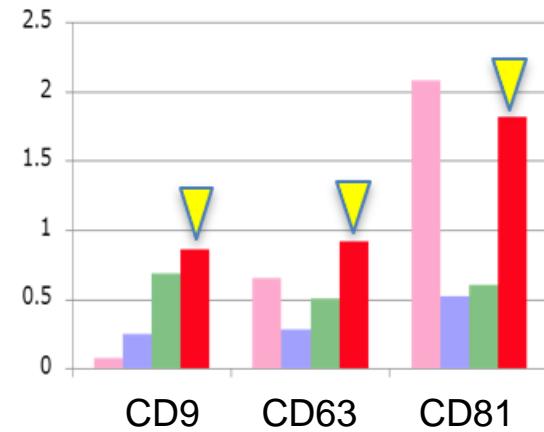
RAJI



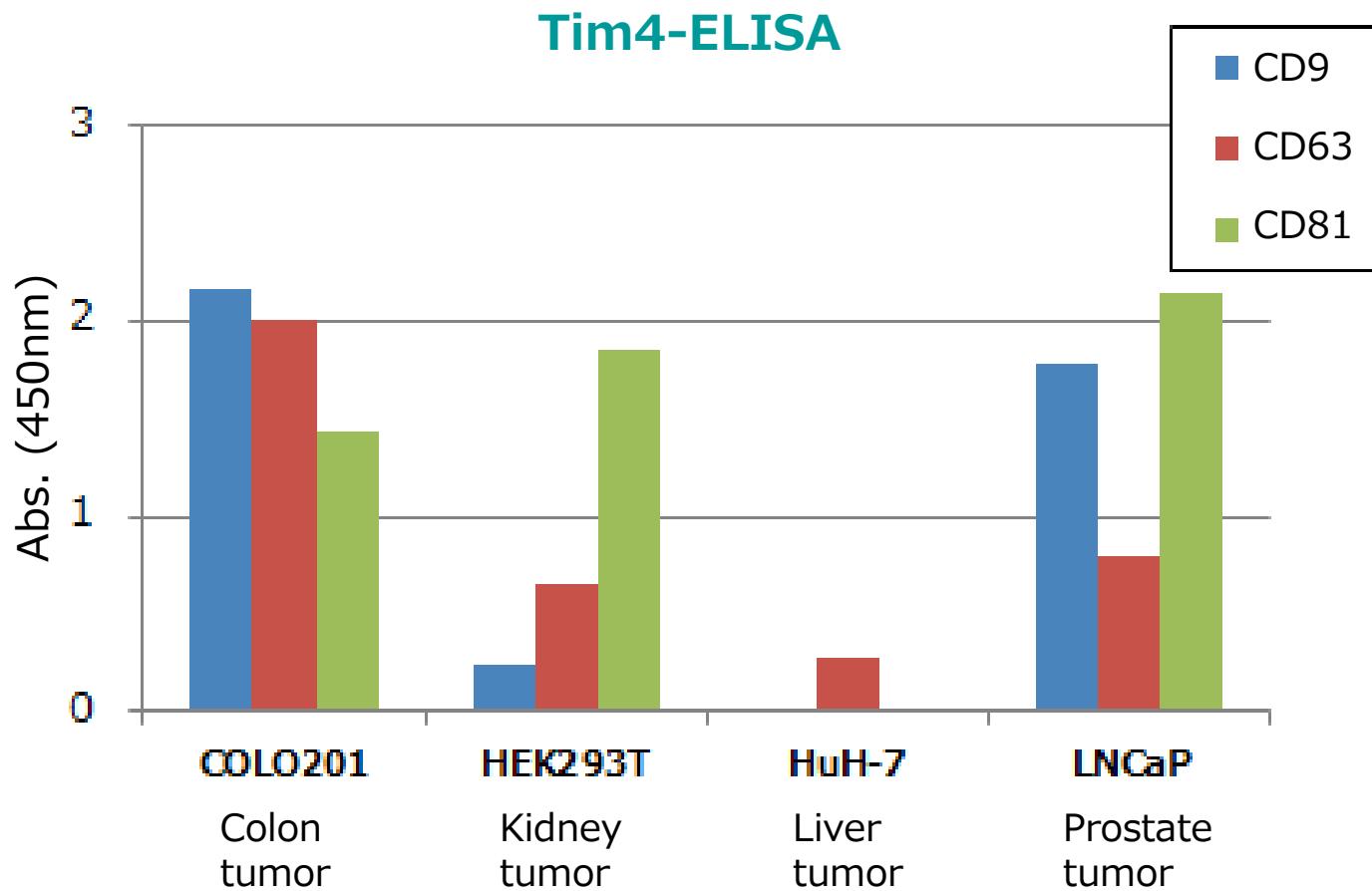
THP1



TIG3



Expression patterns of tetraspanins

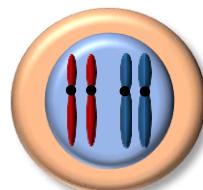


Tim4-ELISA is more sensitive and unbiased method to analyze various sEV subpopulations than using anti-tetraspanin capturing antibodies

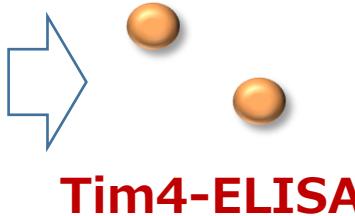
Screening for regulators of sEV secretion

Genetic screening

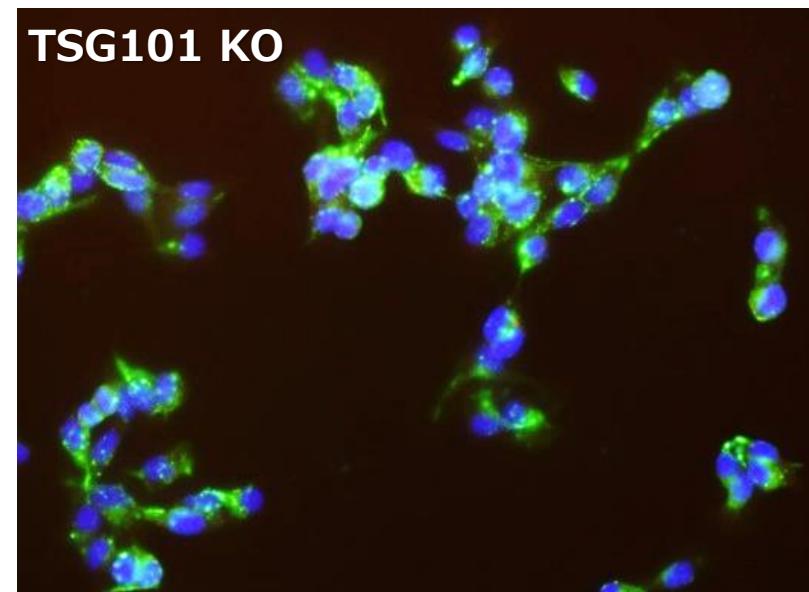
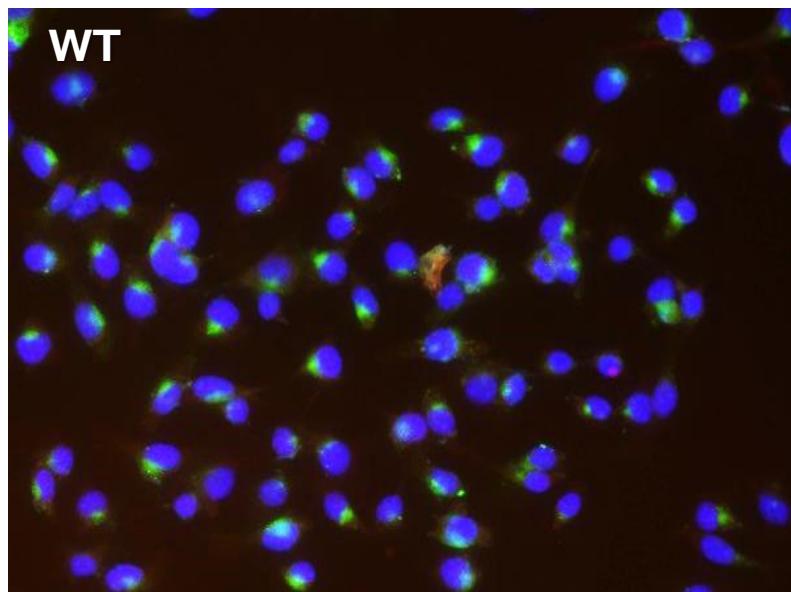
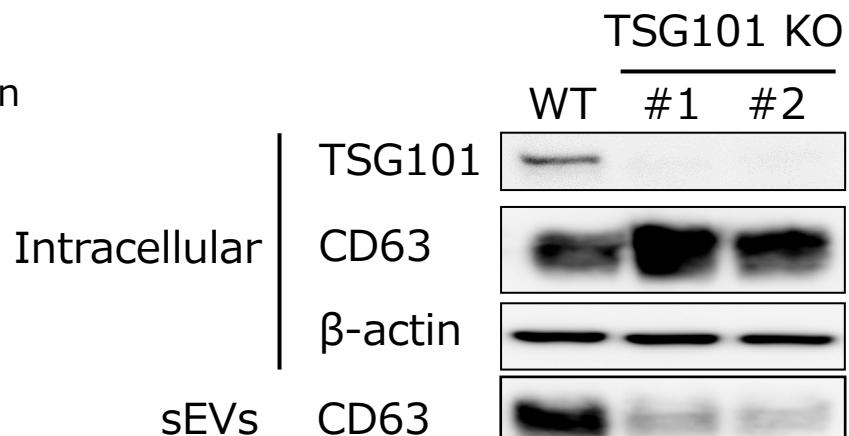
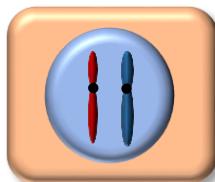
K562 cells
+ shRNA library



Examining changes in
the amount of sEVs

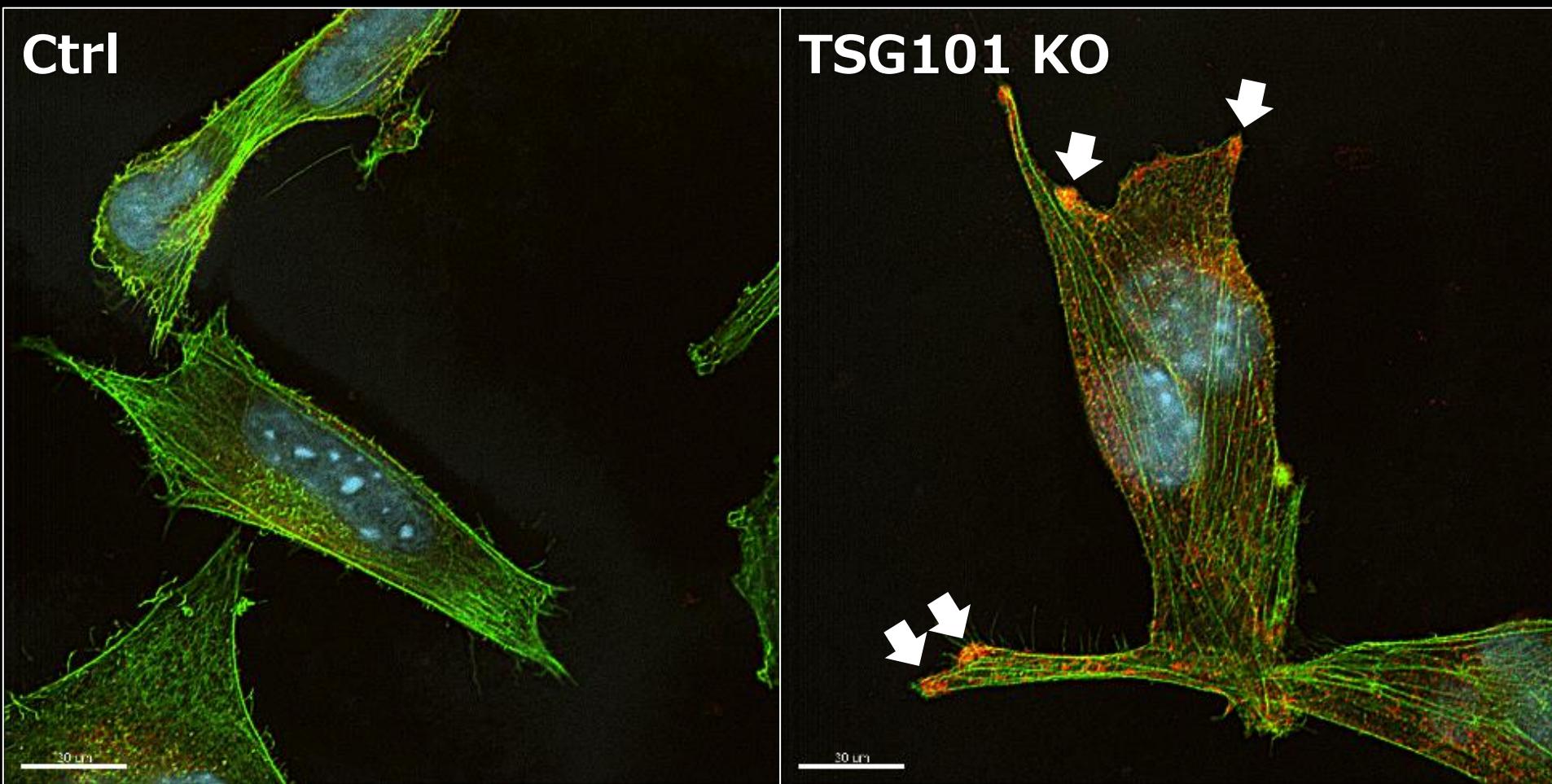


HAP1 cells
+ gene-trap vector



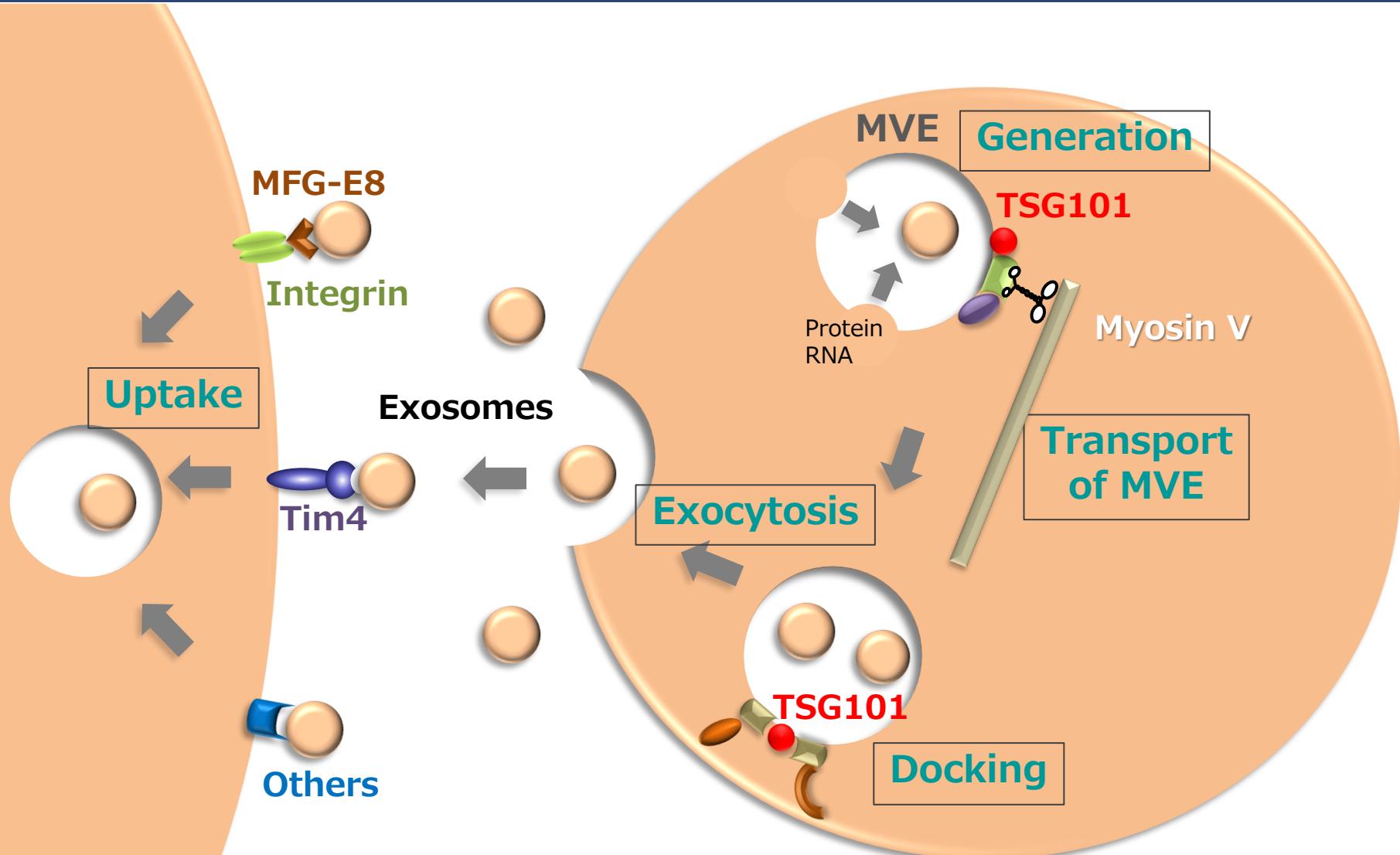
DAPI (Nucleus) / CD63 (Exosomes)

Accumulation of non-secreted vesicles in TSG101-KO



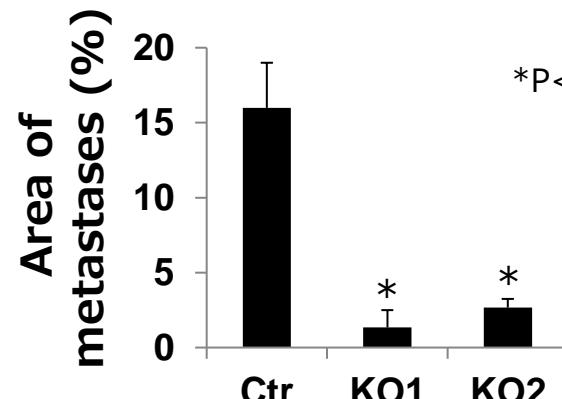
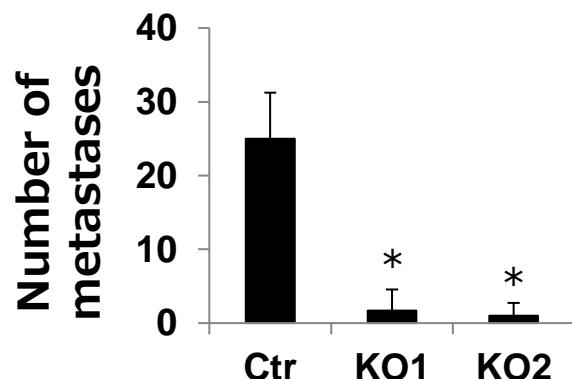
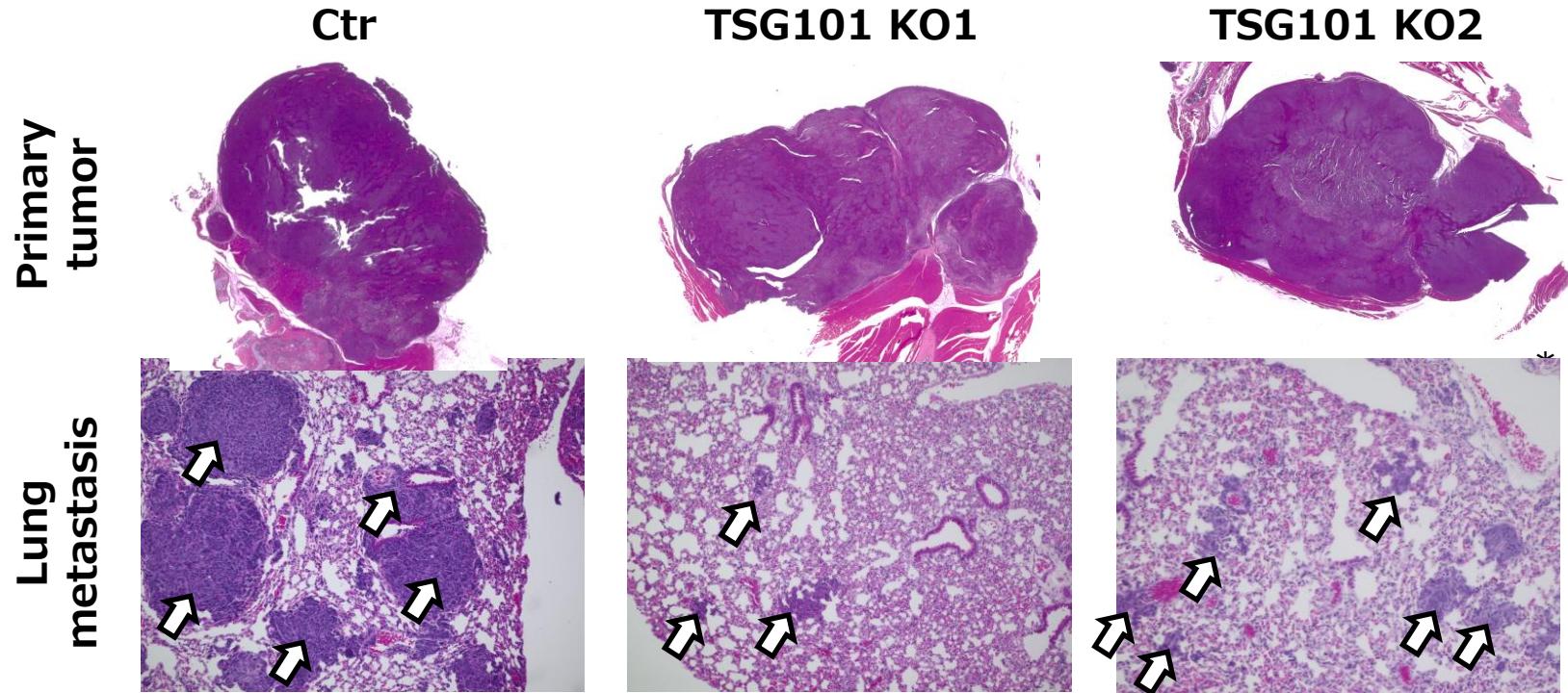
DAPI (Nucleus) Phalloidin (Actin) CD63 (ILVs)
B16F10 cell

A working model for exosome secretion and uptake

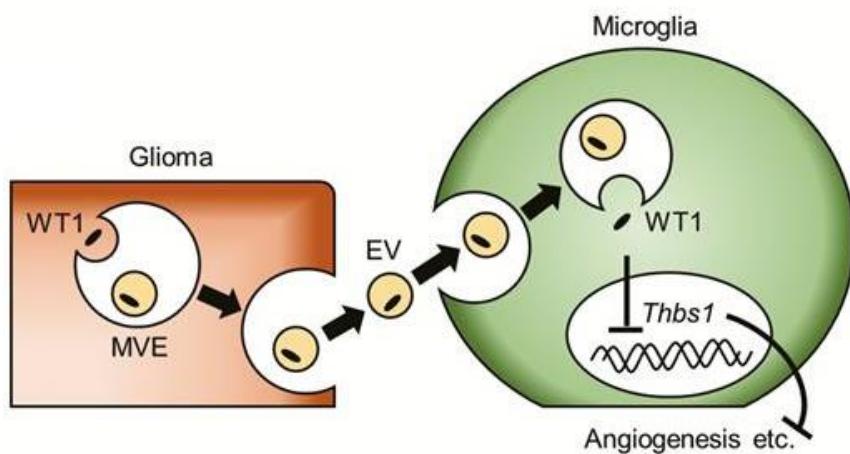
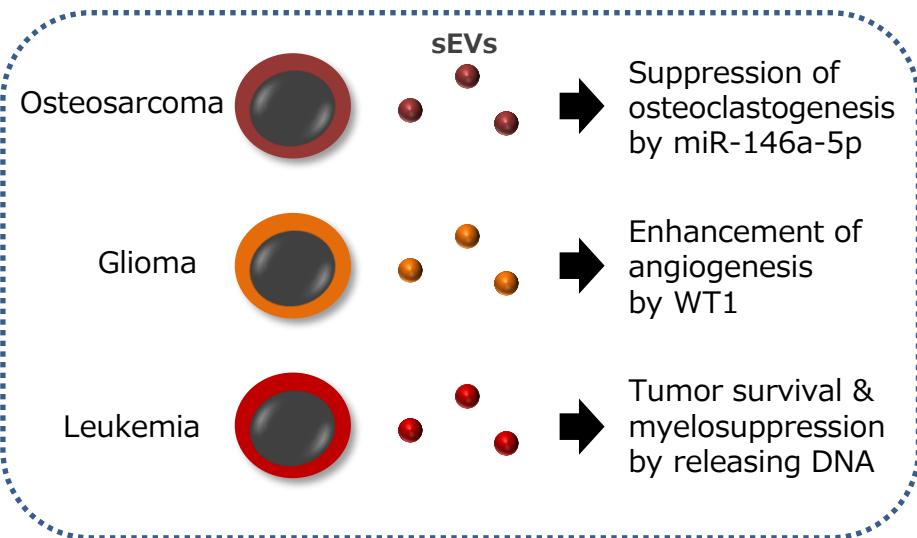


TSG101 KO reduces tumor metastasis

Osteosarcoma



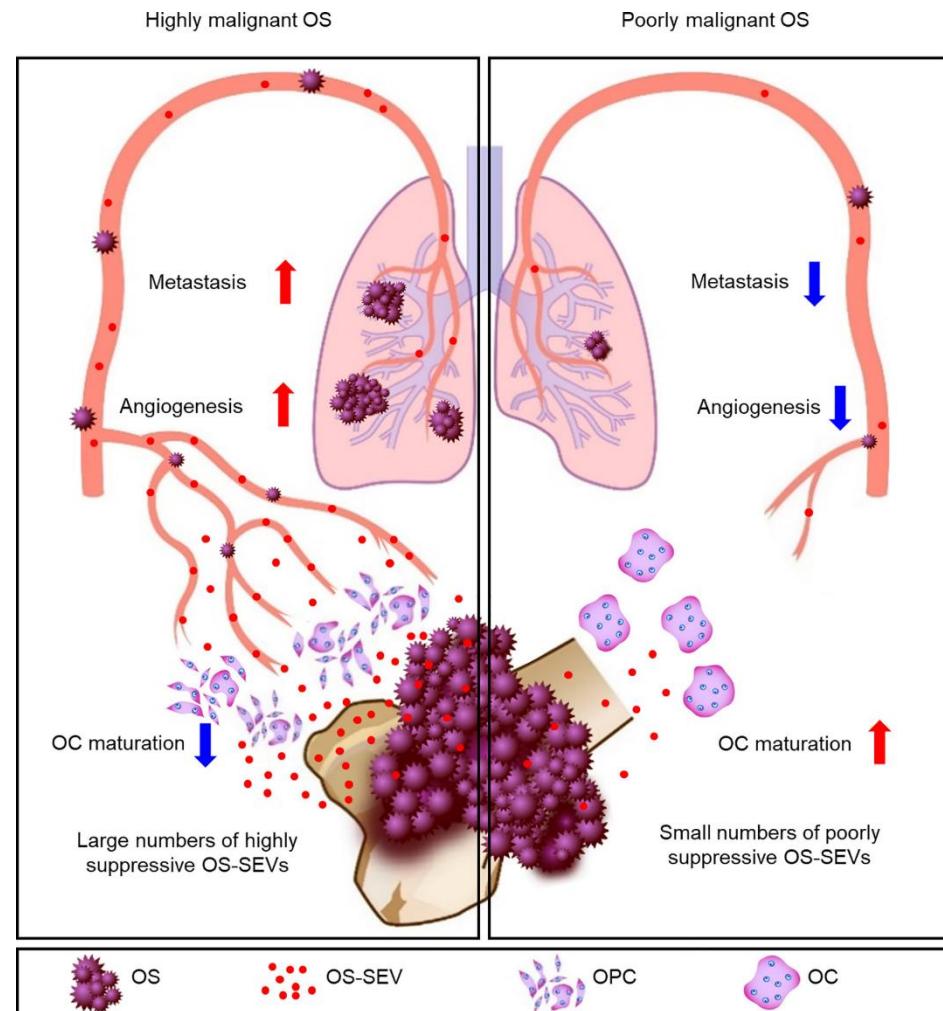
Mechanisms that promote tumor progression by sEVs



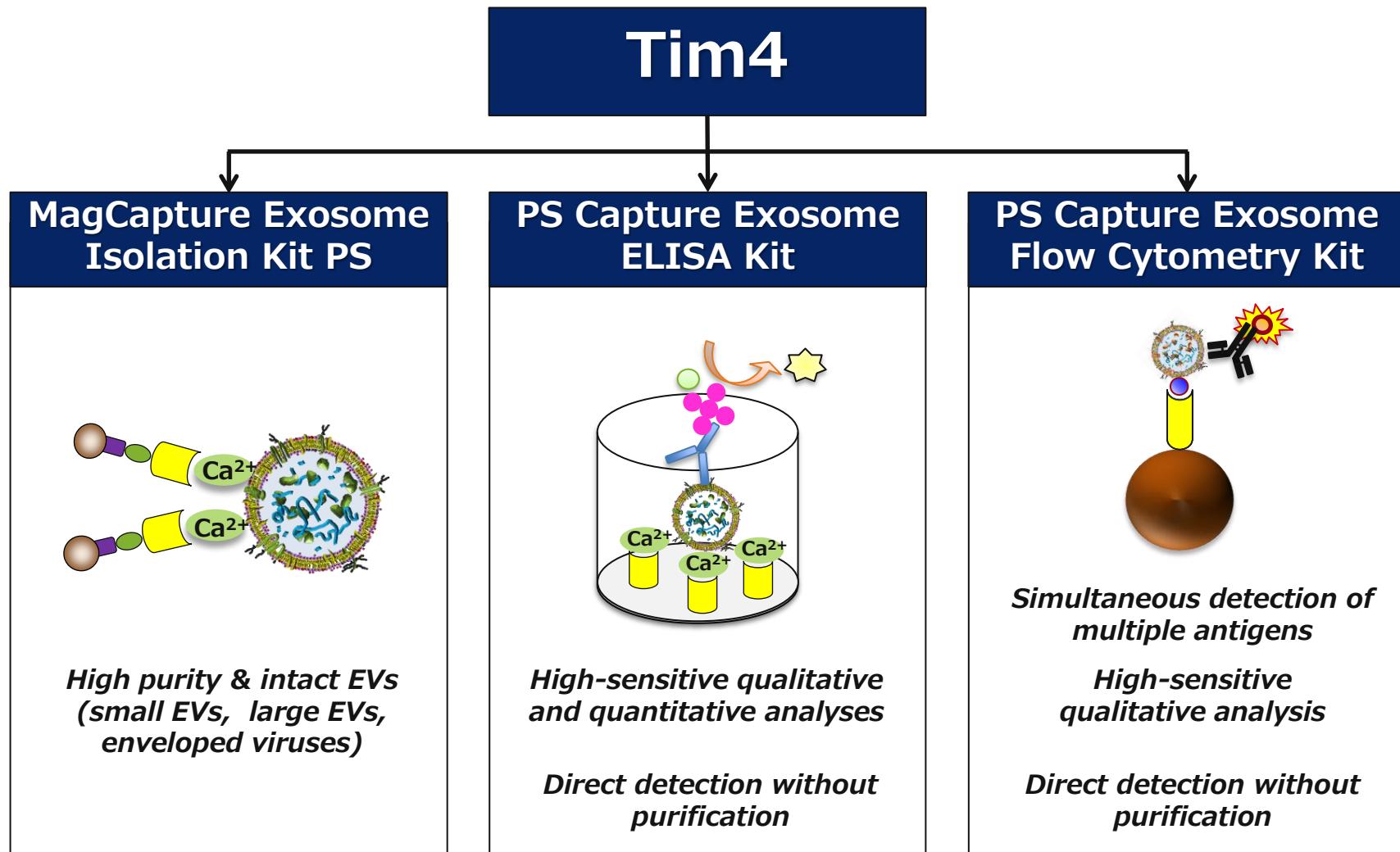
Tsutsui & Hanayama,
Carcinogenesis (2020)

Baba, Hanayama, Mukaida,
Cell Death Dis (2021)

Araki & Hanayama,
Front Oncol (2021)

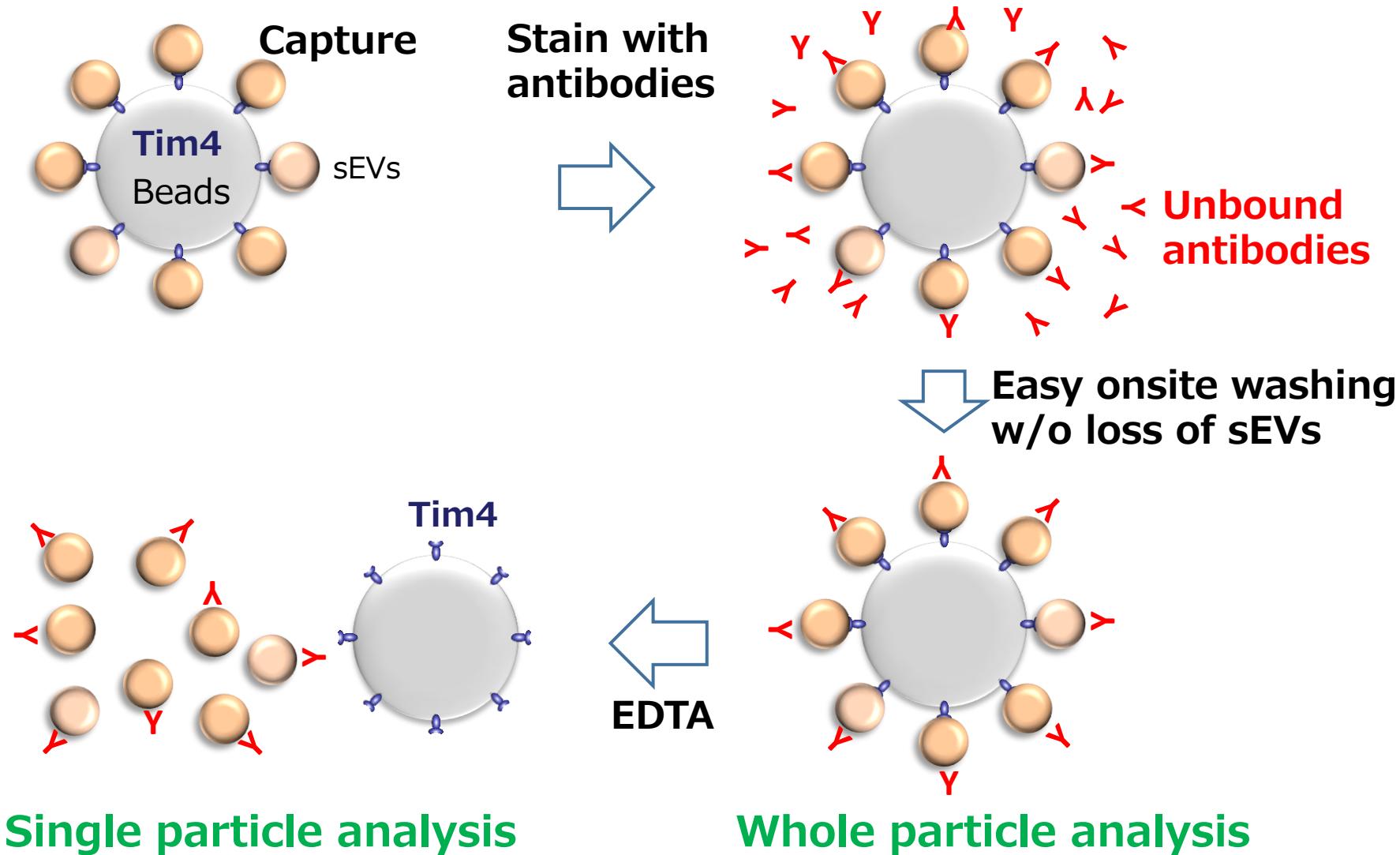


EV research products using Tim4



With easy operation and high reproducibility

Sample preparation for flow cytometry using Tim4



Single particle analyses of sEVs using Tim4-beads



NanoFCM

Tim4
beads

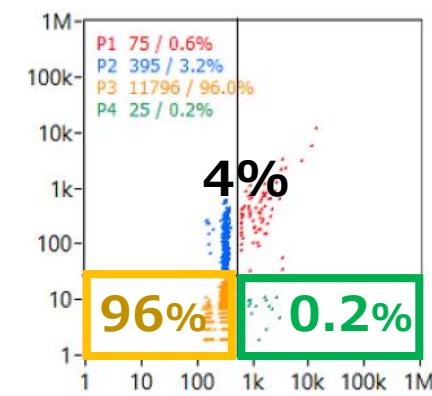
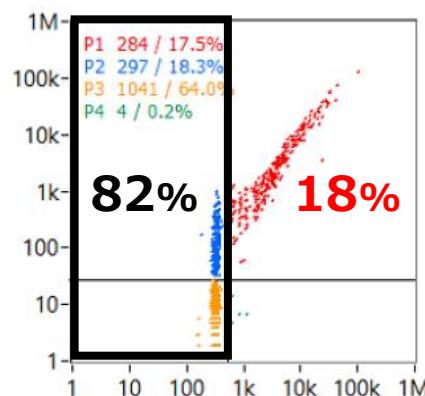
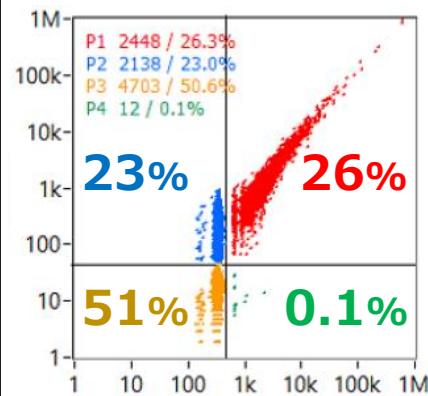
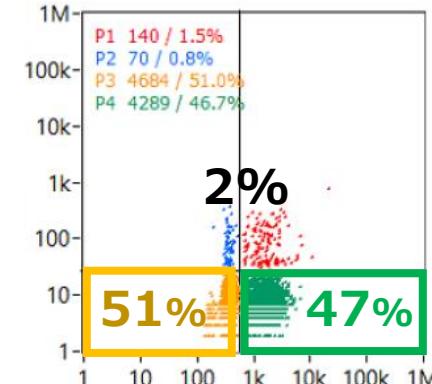
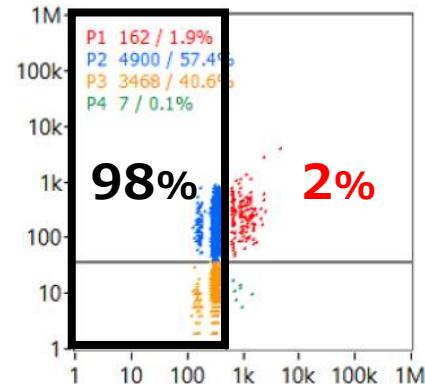
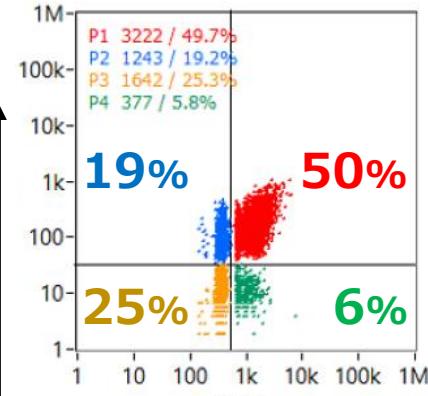
CD9

UC

WT-sEV

CD63KO-sEV

CD9KO-sEV



CD63

Tim4 method is more suitable for sEV sample preparation for FCM analyses

Acknowledgements



FUJIFILM
Value from Innovation

